Research Journal of Pharmacognosy (RJP) 11(1), 2024: 109–136 Received: 22 July 2023 Final revision: 22 Sep 2023 Accepted: 20 Nov 2023 Published online: 26 Nov 2023 DOI: 10.22127/RJP.2023.404256.2145



Matricaria chamomilla: an Updated Review on Biological Activities of the Plant and Constituents

Mina Saeedi^{1, 2}⁽¹⁾, Mahnaz Khanavi³, Kasra Shahsavari⁴, Azadeh Manayi^{2*}⁽¹⁾

¹Persian Medicine and Pharmacy Research Center, Tehran University of Medical Sciences, Tehran, Iran. ²Medicinal Plants Research Center, Faculty of Pharmacy, Tehran University of Medical Sciences, Tehran, Iran.

³Department of Pharmacognosy, Faculty of Pharmacy, Tehran University of Medical Sciences, Tehran, Iran.

⁴School of Medicine, Tehran University of Medical Sciences, Tehran, Iran.

Abstract

Chamomile or camomile (*Matricaria chamomilla* L. syn. *Matricaria recutita* L.) belongs to the family Asteraceae. It is native to Europe and West Asia and has spread to other parts of the world. The plant essential oils and extracts have been frequently used for thousands of years in traditional and folk medicines across the world, due to their valuable medicinal properties. Currently, it is widely applied in different industries such as pharmaceutical, cosmetics, and food industry. Herein, the literature was carefully reviewed via search engines such as Google Scholar, Pub Med, and Scopus using keywords including biological activity, chamomile, flavonoids, pharmacological activity, *Matricaria chamomilla*, and *Matricaria recutita*. Sesquiterpenes such as bisabolol oxide B, bisabolone oxide, and bisabolol oxide A have been identified as the major constituents of chamomile essential oil. Also, various phenolic compounds and flavonoids were mostly reported as active compounds in the plant extracts. Although there are various reports pinpointing the mechanisms of action of chamomile and its constituents, some points have remained ambiguous and further well-designed clinical trials are required. Focusing on the importance of valuable biological properties of chamomile, the present review precisely discussed the characterized chemical constituents of the plant along with their mechanisms of action.

Keywords: biological assays; chamomile; essential oil; Matricaria chamomilla phytochemicals

Citation: Saeedi M, Khanavi M, Shahsavari K, Manayi A. *Matricaria chamomilla*: an updated review on biological activities of the plant and constituents. Res J Pharmacogn. 2024; 11(1): 109–136.

Introduction

Matricaria chamomilla L. synonym *Matricaria recutita* L. belongs to the family Asteraceae, commonly known as chamomile or camomile. It is an annual branching plant with finely divided leaves that reaches the heights of 30 to 60 cm. The plant is native to Europe and West Asia, while it has spread worldwide to the most temperate zones. It has been counted among the medicinal treasures of various culture groups and used in the pharmaceutical and cosmetic

industries with a long history of application in traditional medicine dating back to 7000-9000 BC, referenced by Hippocrates, Galen, and Asclepius [1,2].

Sesquiterpenes are the most dominant compounds identified in the chamomile essential oil. Bisabolol oxide A, bisabolol oxide B, and bisabolone oxide have been frequently reported as the main constituents of the plant essential oil in the literature. However, more than 200

*Corresponding author: manayi@tums.ac.ir

© 2024. Open access. This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (https://creativecommons.org/licenses/by-nc/4.0/)

compounds have been identified in the plant essential oil with different origins. Isolated compounds from the extracts generally include polyphenols, flavonoids like apigenin, luteolin, quercetin, patuletin, isorhamnetin and their derivatives. coumarins, peptides, and polyacetylene [3-5]. It has been well documented that multi-therapeutic properties, cosmetic, and nutritional applications of chamomile both in traditional medicine and current scientific research, are indebted to important classes of volatile and non-volatile active compounds obtained from its essential oils and extracts. Also, different commercial products like tea, infusion, liquid, and capsules have been provided from chamomile [6-8].

According to the studies of both experiments and clinical reports [9], various biological and pharmacological properties including antioxidation, anti-inflammation, antimicrobial, cytoprotection and neuroprotection, wound healing, antiatherogenic and antithrombotic, antidiabetic, gastroprotection, hepatoprotection, antimigraine, and anti-anxiety activities were attributed to the chamomile extracts and essential oils. Moreover, the chamomile extracts and oil market size is noticeable and is expected to grow in the pharmaceutical field as well as perfumery, cosmetics, aromatherapy, and the food industry [10]. In this respect, this review discussed the properties of the plant focused on the identified active constituents and their mechanisms of action to find whether the available data is sufficient.

Methods

The literature was comprehensively searched without time restriction until March 2023 using search engines like Google Scholar, Pub Med, and Scopus by different keywords such as biological activity, chamomile, flavonoids, pharmacological activity, *Matricaria chamomilla*, and *Matricaria recutita*. The search results obtained with the common name of chamomile were refined with the exact mentioned scientific names. All results were extracted and analyzed to construct a correlation between the plant's constituents and biological activity.

Results and Discussion Constituents

Characterization of chamomile essential oil

Analysis of chamomile essential oil samples has been fully achieved in different countries of the world and more than 200 compounds have been reported by various research groups. The major components include thymol (up to 5.8%), β farnesene (0.8-29.8%), β -bisabolene (up to 19.6%), α-bisabolol oxide B (1.3-35.6%), (Z)-αsantolol (0.1-15.9%), α-bisabolone oxide A (0-63.5%), chamazulene (0.2-19.3%), α-bisabolol oxide A (4.9-75.4%), (Z)-ene-yne-dicyclo ether (0.6-10.3%),cis-lanceol (0.1-5.1%),(Z)spiroether (5.1%), cis-pinocamphone (0-73.5%), (Z,Z)-farnesol (1.1-8.3%), and 8-isobutyryloxy isobornyl isobutyrate (11.1-14.0%) [11-17]. The content and composition percent not only depend on the geographical area and climatic conditions of the plant collection site but are also associated with the methods and techniques applied for the preparation of the essential oil, which may include hydro-distillation, supercritical fluid extraction, and headspace analysis.

The study reported by Pirzad et al. [15] showed that the essential oil content and chemical compositions of chamomile were affected by different irrigation regimes (100, 85, 70, and 55% field capacity). It was found that irrigation at 70% field capacity afforded a high amount of essential oil and irrigation at 100% field capacity was not appropriate. Also, the effect of saline irrigation water on the agronomical and phytochemical characteristics of chamomile was investigated by Baghalian et al. It was shown that saline irrigation water had no remarkable effect on oil quantity, both yield, and content of biologically important constituents such as α -bisabolol oxide B, α -bisabolon oxide A, chamazulene, α bisabolol oxide A, α -bisabolol, and trans β farnesene [18].

Homami et al. used different methods for oil extraction including microwave-assisted hydrodistillation and hydro-distillation laboratory scale which yielded 0.08 and 0.06%, respectively. α -Bisabolol oxide A (42.3%), chamazulene (15.1%), α -bisabolol oxide B (9.6%), and (*Z*, *Z*)farnesol (8.1%) were the main constituents by microwave irradiation method. However, using laboratory scale method led to the isolation of *cis*-pinocamphone (73.5%) and α -bisabolol oxide A (7.9%) [17].

Rahmati et al. investigated the effect of nitrogen application (in the form of urea), plant density, and climate conditions on yield and oil constituents of chamomile [19]. The application of urea was found to be very important. The best yields of dry flower and essential oil were related to a plant density of 50 pl.m⁻² with 20 g.m⁻² urea fertilization. The main compounds were α bisabolol oxide A (53.45%), α -bisabolol oxide B (9.90%), α -bisabolone oxide A (5.24%), chamazulene (4.29%), (*Z*)- β -farnesene (2.75%), and spathulenol (0.81%). It should be noted that analysis of the plant essential oil revealed presence of di-(2-ethylhexyl)-phthalate which could be associated with contamination of the oil with plasticizers. The contamination with plasticizers can happen through soil, water, or during processing of the plant [20-22].

Characterization of chamomile extracts

Žlabur et al. investigated the effect of different extraction methods and solvents on the composition of bioactive compounds of the chamomile flower extracts. Extraction methods included conventional methods and ultrasoundassisted extraction [5]. It was found that not only the extraction time of bioactive compounds was remarkably decreased by ultrasound-assisted extraction but also the content of polyphenolic compounds following with antioxidant capacity were significantly higher than conventional methods. Also, the ultrasonic treatment afforded an increase in total acid content. However, other properties such as the density, total soluble solids, and pH value did not show considerable changes. In this respect, the extraction of vitamin C from chamomile flowers was directly affected by the extraction conditions. The highest vitamin C content was obtained using the conventional method with EtOH-water (80:20) at room temperature for 35 min. The ultrasound-assisted

method reduced the amount of vitamin C which probably caused by degradation during sonolysis. Bisabolol oxides: Different bisabolol oxide derivatives (Figure 1) have been identified in the methanol extract of flower heads of German chamomile [23]. The compounds 9hydroxybisabolol oxide A 1, seco-bisabolol oxide B 2, (1R)-l-hydroxybisabololoxide B 3, and bisabolol oxide A glycoside 4 were obtained *n*-hexane fraction while from 15hydroxybisabolol oxide A β -D-glucoside 5 was isolated from the combination of ethyl acetate and *n*-butanol fractions.

Carboxylic acids and esters: The presence of caffeic acid and chlorogenic acid in the chamomile extract [6,24] was reported by Fonseca et al. [24]; (Z)- and (E)-2- β -Dglucopyranosyloxy-4-methoxycinnamic acid [25] were also identified in the ethanol extract of chamomile flowers using HPLC/MS and HPLC/NMR techniques as the precursor of biosynthesis of herniarin in the plant [26]. Moreover, 3-caffeoylquinic acid. 4caffeoylquinic acid, 5-caffeoylquinic acid, quinic acid, ferulic acid-1-O-glycoside, ferulic acid-7-Oglycoside, dicaffeoylquinic acid derivative, 1,3dicaffeoylquinic acid, dicaffeoylquinic acid derivative, quinic acid derivative, caffeoylquinic acid derivative [27], neochlorogenic acid, cryptochlorogenic acid, isochlorogenic acid A, isochlorogenic acid B, isochlorogenic acid C [28], and 1-O-2'-hydroxy-4'-methoxycinnamoyl-β-Dglucose [29] were reported to be isolated from the chamomile extracts.

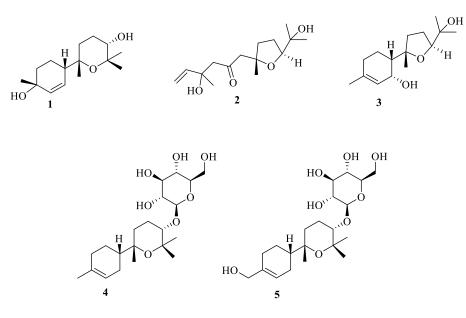


Figure 1. Bisabolol oxide derivatives isolated from *Matricaria chamomilla*

Coumarins: Coumarin (2H-chromen-2-one) [30] and its derivatives (Figure 2) such as herniarin, umbelliferone, aesculetin (cichorigenin), scopoletin and isoscopoletin have been found in the chamomile alcoholic extract. Among them, umbelliferone and herniarin were found to be more abundant [30,31]. Molnar et al. compared different methods such as soxhlet, hydrodistillation, maceration, and supercritical CO₂ extraction, for the isolation of umbelliferone and herniarin in chamomile [32]. The highest yield of umbelliferone and herniarin were obtained using a maceration technique with 50% aqueous ethanol solution. Herniarin and umbelliferone have been identified as stress metabolites. The increased amount of umbelliferone was reported in the leaves of chamomile, 12 h after abiotic stress elicitation by CuCl₂ and this amount

increased approximately 10 times within 48 h. Also, the content of herniarin increased along with a decrease of its precursor, (Z)- and (E)-2- β -D-glucopyranosyloxy-4-methoxy cinnamic acid [33]. Skimmin, daphnin, and daphnetin were found in diploid and tetraploid leaves and anthodia of M. chamomilla L., reported by Petrul'ová-Poracká et al. for the first time [34]. Flavonoids: Flavonoids and their glycoside derivatives are ubiquitous in the extracts of M. chamomilla. According to the Haghi et al. report, the total flavonoid contents of aerial parts of M. chamomilla ranged from 0.82-36.75 g quercetin equivalent (QE)/100 g in dry material [35]. Also, the total phenolic contents of aerial parts of M. chamomilla ranged from 1.77 to 50.75 gram (g) of gallic acid equivalent (GAE)/100 g.

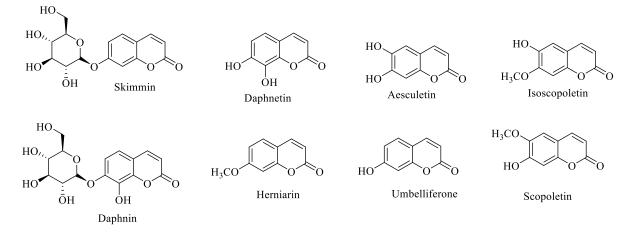
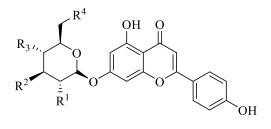


Figure 2. Coumarin derivatives isolated from Matricaria chamomilla



$$\begin{split} & R^{1} = OH, R^{2} = OH, R^{3} = OH, R^{4} = OH \\ & R^{1} = Acetyl, R^{2} = OH, R^{3} = OH, R^{4} = OH \\ & R^{1} = OH, R^{2} = Acetyl, R^{3} = OH, R^{4} = OH \\ & R^{1} = OH, R^{2} = OH, R^{3} = Acetyl, R^{4} = OH \\ & R^{1} = OH, R^{2} = OH, R^{3} = OH, R^{4} = Acetyl \\ & R^{1} = Acetyl, R^{2} = Acetyl, R^{3} = OH, R^{4} = OH \\ & R^{1} = OH, R^{2} = Acetyl, R^{3} = Acetyl, R^{4} = OH \\ & R^{1} = OH, R^{2} = OH, R^{3} = OH, R^{4} = Malonyl \\ & R^{1} = OH, R^{2} = OH, R^{3} = OH, R^{4} = Caffeoyl \\ & R^{1} = OH, R^{2} = OH, R^{3} = Acetyl, R^{4} = Malonyl \\ & R^{1} = OH, R^{2} = OH, R^{3} = Acetyl \\ & R^{1} = OH, R^{2} = OH, R^{3} = Acetyl \\ & R^{1} = OH, R^{2} = OH, R^{3} = Acetyl \\ & R^{1} = OH, R^{2} = OH, R^{3}$$

Figure 3. Flavonoid derivatives isolated from Matricaria chamomilla

The presence of apigenin and its derivatives (apigenin-7-O- β -glucoside, apigenin-7-Opigenin-7-O-glucosyl-6"apiosylglucoside, apigenin-7-O-glucosyl-2"-acetate, acetatee, apigenin-7-Oglucosyl-2",3"-diacetate, and apigenin-7-O-glucosyl-3",4"-diacetate), luteolin and its derivatives (luteolin-7-O-\beta-glucoside, luteolin-4'-O- β -glucoside, luteolin-7-O-βrutinoside, and 6-hydroxyluteolin-7-glucoside), quercetin and its derivatives (quercetin-7-O-Bquercetin-3-Oβ-rutinoside, glucoside, and quercetin-3-O- β -galactoside), patuletin and its (patuletin-7-O-βglucoside), derivative isorhamnetin and its derivative (isorhamnetin-7-O- β -glucoside), chrysoeriol-7-O β -glucoside [36], jaceidin, chrysosplenol, eupatoletin, spinacetin, axillarin, eupalitin, chrysosplenetin, astragalin [37], and naringenin [24] have been reported in the literature. Isorhoifolin (apigenin-7-Orutinoside) as well as various acylated derivatives of apigenin 7-O-glucoside (Figure 3) have been also isolated from the flower of C. recutita [25,38-40].

Peptide: A peptide, MCh-AMP1possessing antifungal activity was purified by reverse phase HPLC from flowers of *M. chamomilla* by Seyedjavadi et al. The confirmed sequence was LSVKAFTGLQLRGVCGLEVKARG [4].

Polyacetylenes: Polyacetylene tonghaosu ((Z)-Ene-yne-dicyclo ether) was identified in the hexane fraction of methanol extract of chamomile flower heads. This compound is chemically unstable and transforms to spirolactone, 1,6-dioxaspiro[4.4] non-3-en-2-one under a photochemical reaction (Figure 4) [3].

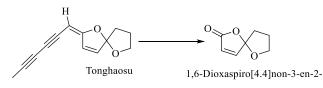


Figure 4. Transformation of tonghaosu to 1, 6 dioxaspiro [4.4] non-3-en-2-one

The mineral elements: Mineral components play an auxiliary role in inducing the biological properties of chamomile. It has been perceived that wild populations contain higher amounts of mineral elements than the cultivated Degumil type. However, the ratios of K/Na and Ca/Mg in the cultivated type are mostly higher than in wild chamomile populations. Mineral content in infusions of chamomile is relatively low in which relatively high concentrations of potassium, calcium, and magnesium were found. The dissolution of mineral elements in tea was reported between 10-26% for most of the elements and the highest value was associated with magnesium (26%) [41].

Chizzola et al. reported the amounts of calcium, potassium, magnesium, zinc, and cadmium as 19-23, 65-73, 47-48, 2324, and 15-21%, respectively, in boiling water extraction of chamomile flower heads within 10-20 min [42]. A longer extraction time led to slightly higher recovery rates for potassium and zinc.

Biological activities of chamomile: in vitro and in vivo and studies

Biological activities of essential oil constituents considering their relevant mechanisms of action are reported in Table 1. Various pharmacological properties of chamomile and its essential oil constituents, in several health conditions related to oxidation, inflammation, microbial infections, and cancer disorders have been fully documented in the literature.

The biological activities of the main constituents of chamomile extracts along with their mechanisms of action are summarized in Table 2.

Antioxidant activity

Numerous chronic pathologies like cancers and cardiovascular diseases are associated with lipid peroxidation which is triggered by free radicals. DPPH, 2.2-Diphenyl-1-picrylhydrazyl, (•) is a stable lipophilic radical that is applied as a model for the evaluation of antioxidant activity in vitro. Methanol extract of chamomile showed higher radical scavenging activity in comparison to other extracts obtained with ethanol, diethyl ether, and hexane which is attributed to the higher phenolic content of the methanol extract including chlorogenic acid, caffeic acid, pcoumaric acid, and ferulic acid [43]. The chamomile flower extract obtained by aqueous ethanol 70% contained a total flavonoid content of 157.9±2.22 mg quercetin equivalent (QE)/g and 21.4±0.327 mg gallic acid equivalent (GAE)/g dry extract and inhibited DPPH with EC₅₀ value of 26.7 µg/mL [44]. A high positive correlation was observed between total phenolics content, total flavonoids, and condensed tannins in methanol and aqueous extracts of chamomile flowers and scavenging of DPPH and 2,2-azinobis-(3-ethylbenzthiazoline-6-sulphonic acid)

(ABTS) radicals [45]. Radical scavenging activity of flower decoction of the plant was significantly higher in comparison to vitamin C against DPPH [46]. Full-fat diet significantly increased SOD, CAT, and GPx in the kidney and liver of rats which were restored to their normal levels by administration of chamomile flower decoction. In addition, the plant decoction increased non-enzymatic antioxidant levels in the kidney and liver [47].

Antimicrobial activity

Essential oil and different extracts of *M. chamomilla* obtained by solvents with different polarities including methanol, ethanol, diethyl ether, and hexane were tested against *Escherichia coli* O157, *Salmonella typhi, Bacillus cereus, Staphylococcus aureus, Candida albicans*, and *Aspergillus flavus* (Table 3). Minimum inhibitory

concentration (MIC) of essential oil generally were in the lower range of 10-12.5 µg/mL in comparison with the plant extracts which inhibited the microorganisms with MIC values ranging from 10-17.5 μ g/mL. The plant EO was rich in α -bisabolol oxide A, α -bisabolol oxide B, and α -bisabolol [43, 48]. α -Bisabolol may inhibit fungal growth through inhibition of ergosterol biosynthesis [49]. Chamomile EO had generally bacteriostatic activity, especially against grampositive bacteria. Chemical composition and biological activities of chamomile extracts were also studied which affected by different solvents with various polarities and methods of extraction (Table 3) [50]. The composition of the plant EO varies based on the season of harvest and method of extraction which can affect the composition and therefore their antibacterial activity [51-53].

Table 1. Biological activities of essential oil constituents of Matricaria chamomilla and their mechanisms of action

Compound	Biological function	Mechanism of action	Ref.
	Anti-inflammatory	In vitro : suppression of NF-κB, p38 and JNK signaling In vivo : increase of the MPO activity and reduction of NO production in systemic infection model in mice; reduction in phosphorylation levels of ERK1/2, p38 and JNK proteins in pulmonary inflammation model; reduction in leukocyte migration, protein extravasations and TNF-α	[129- 132]
	Anti-bacterial	In vitro: inhibition of NorA and TetK efflux pump in Staphylococcus aureus	
α-Bisabolol	Leishmanicidal activity	In vitro: active against <i>Leishmania amazonensis</i> and <i>Leishmania infantum</i> by promotion of programmed cell death process, externalization of phosphatidylserine and membrane damage, reduction in mitochondrial membrane potential and levels of ATP	[134]
	Anti-genotoxic	In vitro: antigenotoxic effect against H ₂ O ₂ by terminating the free radicals	[135]
	Peripheral nervous blocker	In vitro: blocking sodium and potassium voltage-gated channels, decreasing nervous excitability	[136]
	Wound healing	Vound healing In vivo: reduction of NF-κB and elevation of cytokeratin expressions	
	Neuroprotection	In vivo : increase of the MPO activity, TNF-α immunoreactivity in the temporal cortex, and iNOS in both temporal cortex and striatum	[138]
	Nephroprotection	In vivo: nephroprotection in ischemia/reperfusion animal model through antioxidant activity and reduction of tubular damage	[139]
Bisabolol oxide A	Cytoprotective and Cytotoxicity	In vitro: inhibition or delaying necrosis and some apoptotic features through the Ca^{2+} -dependent process (1-10 μ M) while inducing apoptosis and necrosis by concentrations higher than 30 μ M	[140- 142]
	Antipruritic action	In vivo: inhibition of vanilloid receptor activation suppressing sensory irritation and inflammation	[143]
Matricin	Anti-inflammatory	ti-inflammatory In vitro: inhibition of NF-κB transcriptional activity, decrease in adhesion molecule of ICAM-1 and TNF- α	
	Cytotoxicity	In vitro : induction of apoptosis, suppression of cell migration and invasion, caspase activation, and blocking the mTOR/PI3K/AKT signaling pathway	[144]
Chamazulene	Antioxidant	In vitro: scavenging free radicals DPPH and ABTS; inhibition of ROS production	[145,146]
	Anti-inflammatory	In vitro : reduction of TNF-α and IL-6, suppression of the expressions of COX-2, iNOS, MMP-3, MMP-9, and p65 NF-κB; inhibition of production of leukotriene B4, block chemical peroxidation of arachidonic acid	[124,125]
COX 2: evel	In vivo : decrease in the levels of TNF-α and IL-6 ooxygenase: GGT: gamma glutamyl transferase: GSH: glutathione: iNOS: inducible nitric oxide synth		

COX-2: cyclooxygenase; GGT: gamma glutamyl transferase; GSH: glutathione; iNOS; inducible nitric oxide synthase; IFN: interferon; IL: interleukin; JNK; c-Jun N-terminal kinase; MAPK: mitogen activated protein kinase; MMP-9: matrix metalloproteinase-9; MPO: myeloperoxidase; NO: nitric oxide; NF-κB: nuclear factor kappa B; ROS: reactive oxygen specious; SOD: superoxide dismutase; TNF: tumor necrosing factor

Compound	Biological function	tituents of <i>Matricaria chamomilla</i> extracts and their mechanisms of action Mechanism of action	Ref.	
r		In vitro : inhibition of enzymes COX-2 and iNOS, suppression of production of		
		NO, inhibition of pro-inflammatory cytokines like IL-1 β , IL-8, and TNF- α	[100, 100	
	Anti inflommator	secretion, TNF-induced NF-kB transcriptional activation	[120-122 126,147-	
	Anti-inflammatory	In vivo: down-regulation of P2X7/NF-kB pathway, inhibition of IL1b,		
		MMP3,1,13 ADAMTS-5 in arthritis rheumatoid; decrease in inflammatory cell	151]	
		infiltration to synovial		
	Antioxidant	In vitro: DNA protection against free radicals	[152]	
	Anti-mutagenic	In vitro, In vivo, Ames test: prevention or inhibition of genotoxicity	[153-155]	
		In vitro: reduction of α -hemolysin of S. aureus, enhancement of		
		antibiotic activity by inhibition of peptidoglycan synthesis, β -lactamase		
		enzymes, and alteration in permeabilization of outer membrane and		
		cytoplasmic membrane, reduction of Helicobacter pylori colonization and		
	Anti-bacterial	monocyte and neutrophil infiltrations with atrophic gastritis, inhibition of	[156-162]	
		water-insoluble glucans synthesis by Streptococcus mutans, targeting DNA	-	
		gyrase, d-alanine ligase and the type II fatty acid synthetic pathway		
		In vivo: decrease in cytokines in a mouse model of <i>Staphylococcus aureus</i>		
		pneumonia		
		In vitro: inhibition of cytopathogenic effect and replication, interference with		
		viral IRES activity and inhibition of virus induced JNK activation critical for		
	Anti-viral	virus replication EV71, replication inhibition of HCV, interference with the	[163-166]	
	7 thti vitur	translational activity of FMDV by internal ribosome entry site, prevention of	[105-100]	
		cytopathic effect and inhibition of ASFV-specific protein synthesis		
		In vitro : cell shrinkage induction in <i>Candida albicans</i> resulting in alteration of		
	Antifungal	the cell membrane potential and causing leakage of intracellular components	[167]	
Apigenin	Antiparasitic	In vitro: antiproliferation activity against <i>Leishmania amazonesis</i> via rupturing	[169 170]	
	Anuparastuc	of the trans-Golgi network, swelling in parasite mitochondria, alteration in	[168-170]	
		mitochondrial membrane potential, and cytoplasmic vacuolization		
	Antidiabetic	In vivo: reduction of NO, MDA, ICAM, and glucose, increase of GSH, SOD	[171]	
		and insulin		
	Effect in autoimmune diseases	In vivo: in lupus inhibition of APCs and autoantigen presentation, apoptosis of		
		the hyperactive cells, prevention of the production of autoantibodies against	[157,172-	
		nuclear antigens; in MS, reduction of T cells proliferation, restrain		
		phagocytosis, producing IFN-y, ROS, regulation of miR-155 in inflammation;		
		in myocarditis suppression of the infiltration of inflammatory cells, modulation	180]	
		of the Th1/Th2 cytokine balance; in ulcerative colitis suppression of the		
		infiltration of inflammatory cells and cytokine production, reduction of MPO,		
		MDA, COX and iNOS		
		In vitro: induction of apoptosis; modulation of glycolytic and mitochondrial		
		pathways of ATP production, blocking IL-6 and IL-10 secretion and up		
		regulation of mRNA expression of TNF- α , caspase 3 and 8; decrease in cancer		
	Cytotoxicity	cell growth, glucose utilization, invasion into matrix, reduction in angiogenesis	[135,181-	
		and endothelial attachment; inhibition of ornithine decarboxylase; down	184]	
		regulation of MMP-2, -9 and slug and snail, down regulation of NF-KB		
		p105/p50, PI3K, and Akt and its phosphorylation; activation of proteasomal		
		degradation apparatus		
	Summassian of	In vivo: suppression of metastasis in animal models; delay in tumor growth;	[107 10F	
	Suppression of	suppression of IL-6-linked downstream signaling pathway resulting in anti-	[182,185-	
	cancer	invasive effect	189]	
	Neuroprotection	In vivo: providing anti-ischemic effect through increase in activity of HO-1	[100]	
	Neuroprotection	and Nrf2	[190]	
	Anti-inflammatory	In vitro : inhibition of TNF- α secretion	[120]	
		In vitro: augmentation of the homeostatic potential of HO-1 through		
		modulation of the Nrf2/MAPK signaling pathway; inhibition of Ang II-		
		mediated oxidative stress and apoptosis in macrophages; antagonize p38		
	Antiatherogenic	MAPK apoptotic signal		
	property	In vivo: inhibition of proatherogenic mediators like IL-1, TNF-α, and MMP-9,	[191-197]	
		ICAM-1, VCAM-1, and E-selectin in LPS-challenged mice; suppression of		
		serum TG, total cholesterol, LDL, creatine kinase, lactate dehydrogenase, and		
Luteolin				
Luteolin		CTGF in male rats; attenuation of oxidative stress and dysfunction of eNOS in		
Luteolin		CTGF in male rats; attenuation of oxidative stress and dysfunction of eNOS in		
Luteolin		CTGF in male rats; attenuation of oxidative stress and dysfunction of eNOS in diabetic rats with ischemia and reperfusion injury		
Luteolin	Antithrombotic	CTGF in male rats; attenuation of oxidative stress and dysfunction of eNOS in diabetic rats with ischemia and reperfusion injury In vitro: reduction in fibrin clot mass through suppression of fibrin polymers,		
Luteolin	Antithrombotic	CTGF in male rats; attenuation of oxidative stress and dysfunction of eNOS in diabetic rats with ischemia and reperfusion injury In vitro: reduction in fibrin clot mass through suppression of fibrin polymers, inhibition of enzymatic activities of thrombin and FXa	[198,199]	
Luteolin	Antithrombotic property	CTGF in male rats; attenuation of oxidative stress and dysfunction of eNOS in diabetic rats with ischemia and reperfusion injury In vitro: reduction in fibrin clot mass through suppression of fibrin polymers,	[198,199]	

		hyperglycemia-induced cytokine production through NF-KB suppression	
Quercetin	Anti-inflammatory	In vitro : inhibition of TNF- α secretion	[120]
	Antiatherogenic	 In vitro: inhibition of thrombin, ADP-induced aggregation of platelets, and 12- HETE, reduction of oxidation and aggregation of LDL In vivo: elevation of cytochrome P450 content in the liver of experimental animals, attenuation of lipid peroxidation and hyperlipidemia in the aorta of high cholesterol-fed white rabbit, reduction of LDL oxidation in LDL receptor– deficient mice 	[201-207]
	Suppression of cancer	In vivo: suppression of metastasis in animal models, delay in tumor growth and raising survival rate	[185,187]
Umbelliferone	Antioxidant	In vitro: scavenging DPPH free radical, suppression of membrane reactive free t hydroxyl radical, chelating ferrous ion, inhibition of lipid peroxidation, suppression of ROS production	
	Anti-inflammatory effect		
	Anti-genotoxic	In vitro: inhibition against oxidative DNA damage	[210]
	Antinociceptive	In vivo: antinociceptive property involved in NO system not opioid pathway, inhibition of both peripheral and centrally acting pain mediators	
	Effect on hyperpigmentation	In vitro: weak inhibition of tyrosinase compared to kojic acid	[213]
	Anti-diabetic effect	 In vitro: inhibition of aldose reductase and α-glucosidase; activation of PPARγ and PPARβ In vivo: improvement of glucose tolerance, modulation of hepatic lipid metabolism and antioxidant defense system with increase in levels of adiponectin; protective effect in liver cells by reduction of GGT, AST, and ALT 	[214-217]
	Anti-cancer activity	In vitro: reduction in cell viability and cell migration; induction of cell cycle arrest, apoptosis, and DNA fragmentation	[218-220]
Herniarin	Anti-genotoxic	In vitro: inhibition against oxidative DNA damage In vivo: decrease in the apoptotic and necrotic cells and the ROS level in bone marrow cells	[210,221]
	Cytotoxicity	In vitro: increase in chromatin condensation	[222]

ASFV: African swine fever virus; ALT: alanine aminotransferase; AST: aspartate aminotransferase; COX-2: cyclooxygenase-2; eNOS: endothelial nitric oxide synthase; EV71: enterovirus 71; FMDV; foot-and-mouth disease virus; GGT: gamma glutamyl transferase; GSH: glutathione; HO-1: heme oxygenase-1; HCV: hepatitis C virus; 12-HETE: 12-hydroxyeicosatetraenoate; ICAM: intracellular cell adhesion molecule; iNOS: inducible nitric oxide synthase; IFN: interferon; IL: interleukin; JNK: c-Jun N-terminal kinase; LDL: low density lipoprotein; MAPK: mitogen activated protein kinase; MMP-9: matrix metalloproteinase-9; HO-1: heme oxygenase-1; 5-LOX: 5-lipoxygenase; CTGF: myocardial connective tissue growth factor; MS: multiple sclerosis; MPO: myeloperoxidase; NO: nitric oxide; NF-κB: nuclear factor kappa B; PPAR: peroxisome proliferated activated receptor; Nrf2: E2-related factor 2; ROS: reactive oxygen specious; SOD: superoxide dismutase; TG: triglyceride; TNF: tumor necrosing factor; VCAM: vascular cell adhesion molecule

No growth inhibition was observed by the essential oil of chamomile flower against Salmonella paratyphi, Salmonella typhimurium, Shigella spp., P. aeruginosa, S. aureus, and E. coli which can be explained by the low concentration of a-bisabolol (4%) that known as a responsible component for antibacterial activity [54]. The plant essential oilwas examined against 12 strains of P. aeruginosa and 8 strains of S. aureus which were isolated from patients diagnosed with external otitis. No inhibition was observed against P. aeruginosa, while the plant oil showed inhibition activity against growth of three S. aureus with inhibition zone diameter of 10 mm [55]. While, in another study the methanol extract of chamomile providing inhibition activity against P. aeruginosa with MIC value of 78 µg/mL. The presence of terpenoids, phenols, tannins, and flavonoids were

proven in the extract which may responsible for the observed antibacterial activity [56].

The result of a study showed that hydroethanolic chamomile exhibited extract of small antiadhesive activity in human colon carcinoma cells affected by Campylobacter jejuni [57]. Alternatively, the hydroethanolic extract of the plant was tested against two other responsible bacteria for gastrointestinal diseases. Helicobacter pylori and C. jejuni, showing very satisfactory results against C. jejuni in that with a concentration of 20 mg/mL the growth of 100% inhibited [58]. The bacteria 50% hydroalcoholic extracts of chamomile flowers and leaves were fractioned and tested against Bacilus subtilis and Pseudomonas syringae. Both flowers and leaves suppressed the growth of Pseudomonas syringae and some of the fractions

showed antibacterial activity which was ascribable to *cis*-, *trans*-spiroethers, and the coumarins like herniarin and umbelliferone [59]. The 70% ethanol extract of the flowering part of *M. chamomilla* was tested against *Staphylococcus aureus*, methicillin-resistant *S. aureus* (MRSA), *S. epidermidis*, *P. syringae* pv. *maculicola*, *Xanthomonas vesicatoria*, *Aliivibrio fischeri*, and *Bacillus subtilis*.

The chamomile extract suppressed the growth of *Staphylococcus epidermidis*, MRSA, and *S. aureus* with an inhibition zone diameter ranging 6-7 mm, while vancomycin only inhibited the growth of MRSA with a zone diameter of 7 mm. Two active compounds, apigenin and α -linolenic acid, were detected in the tested extract [60].

Antiparasitic activity

Amoebae of Acanthamoeba castellani was sensitive to α -bisabolol, the main sesquiterpene in the chamomile essential oil, in a dosedependent base with respective IC₅₀ and IC₉₀ values of 20.83 and 46.60 µg/mL. The compound externalization caused apoptosis by of phosphatidylserine, cellular membrane damage, decrease in the mitochondrial membrane potential, and reduction of ATP level [61]. With concentrations of 1000 and 500 μ g/mL α bisabolol, caused 100% inhibition against Leishmania infantum promastigote [62]. Solid lipid nanoparticles containing α -bisabolol with diminazene aceturate in Trypanosoma evansi infected mice improved the therapeutic effectiveness of diminazene aceturate [63].

Plant extract/EO	Microorganisms	MIC (µg/mL)	Ref.
	Escherichia coli	15	
	Salmonella typhi	15	
Methanol extract	Bacillus cereus	12.5	-
Methanol extract	Staphylococcus aureus	12.5	
	Aspergillus flavus	12.5	_
	Candida albicans	10	
	Escherichia coli	15	
	Salmonella typhi	15	
Ethen all and us at	Bacillus cereus	12.5	
Ethanol extract	Staphylococcus aureus	12.5	
	Aspergillus flavus	15	
	Candida albicans	12.5	_
	Escherichia coli	17.5	_
	Salmonella typhi	15	[43]
	Bacillus cereus	15	
Diethyl ether extract	Staphylococcus aureus	15	
	Aspergillus flavus	17.5	
	Candida albicans	15	
	Escherichia coli	15	
	Salmonella typhi	15	
	Bacillus cereus	12.5	
Hexane extract	Staphylococcus aureus	10	
	Aspergillus flavus	12.5	
	Candida albicans	10	
	Escherichia coli	12.5	
	Salmonella typhi	12.5	
	Bacillus cereus	10	
Essential oil	Staphylococcus aureus	10	
	Aspergillus flavus	12.5	
	Candida albicans	10	_
	Staphylococcus aureus	0.01	[48]
	Bacillus cereus	0.02	
	Bacillus subtilis	0.03	
Essential oil	Shigella shiga	0.14	
	Shigella sonnei	0.2	
	Pseudomonas aeruginosa	4	_
	Proteus sp.	0.15	_
Ethanol extract	Pseudomonas aeruginosa	106	[223]

Table 3. Antimicrobial activity of Matricaria chamomilla extracts and essential oil against different microorganisms

MIC: minimum inhibitory concentration; EO: essential oil

The ethyl acetate fraction of chamomile showed the highest antioxidant activity with stronger amoebicidal activity against *Acanthamoeba castellanii* in comparison with other fractions of extract including hexane, chloroform, and aqueous fractions. The analysis of the ethyl acetate fraction indicated the presence of a mix of coumarin derivatives in it [64].

Food preservation

The application of chamomile as a food preservative was evaluated in some studies. chamomile improved Decoction of the antioxidant potential of cottage cheese by lowering lipid peroxidation and the higher free radical scavenging activity that prolonged the shelf life of the cheese. This bioactivity may be related to the release of phenolic compounds possessing antioxidant and antimicrobial activities [65]. The microencapsulation of the extract also prolonged antioxidant activity in the cheese after seven days, but had similar color parameters and nutritional as well as fatty acid profiles [66]. Decoction of chamomile as an antioxidant agent was used in wheat biscuits, and samples were analyzed for 60 days of storage monitoring nutritional profile, sugars, fatty acids, and antioxidant capacity. The incorporation of natural or synthetic antioxidants did not significantly change the color or nutritional values of the biscuits. Similar antioxidant capacity was achieved by both antioxidants; therefore, the extract of the plant is a good replacer for synthetic additives [67]. Matricaria chamomilla essential oil was applied in caseinbased film to increase its barrier properties and antibacterial activity. The film showed an inhibitory effect against Listeria monocytogenes, S. aureus, and E. coli in vitro which was evaluated by the disk-diffusion method. α -Bisabolol and other components in the oil were responsible for the antibacterial activity of refined casein-based film [68]. The majority of essential oils are classified as generally recognized as safe (GRAS) including M. chamomila. While their application may be limited due to the flavor consideration because doses of the antimicrobial amount of oil may exceed organoleptically acceptable levels [69].

Anthelmintic activity

Methanol and aqueous extracts of flowers of M.

chamomilla which were rich in phenolic compounds, flavonoids, and condensed tannins showed high inhibitory effects on egg hatching. The methanol extract at a concentration of 4 mg/mL inhibited 100% of egg hatching with an IC₅₀ value of 1.52 mg/mL. A high correlation was found between the inhibition of egg hatching and the phytochemical contents of the extracts. After 8 h, the methanol extract (8 mg/mL) induced 91.77% mortality against adult worm Haemonchus contortus, while aqueous extract with the same concentration caused 75.05% mortality. Albendazole as a reference, caused 82.26% mortality [45].

Anti-inflammatory activity

Results of a study showed that hydrophilic Carbopol gel containing ethanol extract of flowers of chamomile was not active in the carrageenan paw edema model in rats. Sodium lauryl sulfate (SLS) was applied as the permeation enhancer in gelling formulations. The presence of apigenin was assessed in the plant extract using the HPLC method [70]. Previous studies revealed that luteolin, apigenin, and chamazulene were individually efficacious in inhibition of COX-2, iNOS, and leukotriene expression in cell culture [71].

Anti-platelet activity

Polysaccharide-polyphenolic conjugates isolated from *M. chamomilla* resulted in a dose-dependent decrease of platelet aggregation induced with Adenosindiphosphat (ADP), collagen, and arachidonic acid. The extract showed no toxicity against human blood platelets, mouse fibroblast L-929, and human lung cells A-549. Therefore, compounds obtained from chamomile have the potential for the development of new anti-platelet agents [72].

Cytotoxicity

The chamomile flower extract, obtained by aqueous ethanol 70%, rich in phenolics, and flavonoids showed cytotoxicity against human hepatoma cancer cells (HepG-2) in a dose-dependent manner with an IC₅₀ value of 300 μ g/mL [44]. Extract of flowering aerial parts of the plant inhibited nitric oxide (NO) production in colorectal cancer cell line HT-29 and exhibited cytotoxicity against the cancer cell line with IC₅₀ values of 1881 and 1669 μ g/mL after 24 and 48 h, respectively [73].

Antiangiogenic activity

Vascular endothelial growth factor (VEGF) stimulates the process of blood vessel formation; expression of VEGF was inhibited by flower extract of *M. chamomilla*. Additionally, the extract of chamomile flower down-regulated matrix metalloproteinase (MMP-9) substantiating the activity that is responsible for the regulation of tumor angiogenesis [44].

Gastroprotective effect

Decoction of chamomile flowers protected gastric mucosal damage caused by ethanol in rats significantly and dose-dependently. Pre-treatment with the flower decoction with a dose of 100 mg/kg reduced gastric lesions by 90.95%, while famotidine or ascorbic acid decreased lesions by 81.13% and 73.12%, respectively. Considering the mechanism of action of the plant, the decoction might partly reverse the depletion of antioxidant enzymes and sulfhydryls induced by ethanol in a dose-dependent manner. Levels of intracellular mediators including H₂O₂, iron, and calcium were also decreased by chamomile significantly. The presence decoction, of phytochemicals like polyphenols, flavonoids, and condensed tannins with high concentrations has been shown to be responsible for the antioxidant activity of the plant [74].

Anti-diarrheal activity

Decoctions of M. chamomilla flower with the doses of 25, 50, and 100 mg/kg were administered in an animal model of diarrhea induced by castor oil significantly reduced the number of defecations. Loperamide with the dose of 20 mg/kg produced a more marked antidiarrheal effect but less than the highest dose of the flower decoction. Intestinal lipoperoxidation with H₂O₂ and labile iron induced by castor oil were reduced with a decoction of chamomile flower in a dosedependent manner. Castor oil depleted mucosa antioxidant enzymes, the effect which decreased with pre-treatment of chamomile decoction [75].

Hepatoprotective activity

The decoction of *M. chamomilla* flower protected liver injury and histological changes dosedependently induced with acute alcohol intoxication in rats. Pretreatment with extract significantly reversed lipoperoxidation, antioxidant enzyme depletion, and attenuated liver-SH group level. Alcohol treatment increased calcium, free iron, and H_2O_2 levels in plasma and liver, while pretreatment with a decoction of chamomile flower showed protection against ethanol-induced intracellular mediator disturbances in a dose-dependent manner [46]. High-fat diet in rats increases activities of ALT and AST which is attenuated with the administration of chamomile flower decoction [47].

Spasmolytic activity

The anti-spasmodic and spasmolytic effects of flavonoid components of M. chamomilla were evaluated on isolated jejunum of rabbits. The plant showed spasmolytic effect on K⁺-induced spontaneous contractions, suggesting that calcium channels are involved in its spasmolytic actions. The results of the study revealed that the spasmolytic effect of the plant was mediated NO-activated cholinergic through and histaminergic receptors [76]. Several other studies indicated the indirect Ca2+ channelblocking activity of chamomile and its flavonoid fraction [77-79]. However, verapamil could not prevent the antispasmodic activity of the plant. The extract of the plant in low doses could augment the effect of verapamil which was irrespective of indirect reaction with Ca²⁺ channels. In addition, the spasmolytic effect of chamomile could be blocked by atropine and cetirizine which possibly indicated that muscle relaxation by chamomile was partly mediated through acetylcholine (Ach) and histidine (His) receptors [80]. Results of a study showed that the spasmolytic effect of M. chamomilla and its flavonoid fraction was mediated by the inhibition of cAMP activity and cAMP phosphodiesterase [81,82]. A growing body of evidence suggests that a wide range of signaling cascades are activated by apigenin and quercetin causing smooth muscle relaxation [83,84]. Apigenin has also been attributed to the inhibition of Ca²⁺ release or induction of eNOS and NO production in mouse gastric tissue. An increase in intracellular calcium concentration by apigenin and quercetin resulted in significant NO production. Thus, smooth muscle relaxation is mediated by the blockade of calcium release and NO production by the flavonoid fraction of chamomile [76,79].

Fibrinolytic activity

In a model of alveolar bone resorption (ABR) in

ethanol extract of M. chamomilla rats. significantly reduced ABR and furcation lesions besides preservation of trabecular alveolar bone cementum. Leukocyte infiltration of and periodontium and MPO activity were also reduced by the plant extract. Levels of TNF- α and IL-1 β with immunostaining for RANKL and TRAP were decreased in the animals treated with the extract. Thus, inflammation and ABR in animals were restricted by reducing neutrophils, TNF- α and IL-1 β , and osteoclastogenesis-related molecules (RANKL/TRAP) or increasing molecules that inhibit the RANK-RANKL interaction (OPG), without interfering with bone anabolism [85].

Antidiabetic activity

Aqueous extract of chamomile enhanced insulin levels and reduced amylase activity and HbA1c in alloxan-induced diabetic rats after six weeks. The plant extract with the doses of 150 and 300 mg/kg reduced fasting plasma glucose by 20% and 57%, respectively [86]. Chamomile extract (200 mg/kg) with physical exercise increased insulin levels and decreased fasting plasma glucose in type 1 diabetic rats for 8 weeks [87]. However, results of another study suggested that extract of chamomile flower (500 mg/kg) showed considerable reduction in serum level of glucose up to 64% without an effect on insulin level in streptozotocin (STZ) diabetic rats model after four weeks [88]. Level of insulin and insulin sensitivity increased in STZ-nicotinamideinduced diabetic male rats by administration of 200 mg/kg/day of chamomile extract for 24 weeks. A significant reduction in fasting plasma glucose and serum GPLD1 levels was also observed in the treatment group in comparison to the control group, while serum levels of glypican-4 remained unchanged [89]. Decoction of chamomile (500 mg/kg) in STZ-induced inhibited diabetic rats hepatic glycogen phosphorylase leading to a significant reduction plasma glucose after 21 days in [90]. Administration of aerial parts of the ethanol extract of chamomile to STZ-induced diabetic rats caused an increase in insulin-positive β -cells of pancreatic islets and decreased glucose levels in the serum of animals dose-dependently [91]. Oral administration of both single dose and a daily oral dose of aqueous extract of chamomile leaf and flower (20 mg/kg) significantly attenuated postprandial blood glucose (61%) in

STZ-induced rats; however, difference in plasma insulin level was not significant [92]. Levels of HbA1c with fasting and post-prandial blood glucose versus baseline reduced in alloxaninduced diabetic rats by administration of chamomile flower tea for 8 weeks [93]. The addition of chamomile decoction to the mucosal reservoir significantly and dose-dependently inhibited intestinal glucose absorption [47]. Aqueous extract of chamomile leaves (300 mg/kg) restored levels of urea, uric acid, creatinine, and total protein to normal levels in diabetic rats. The plant extract also maintained the weight gain and reduced apoptosis in the renal cells by reversing lower level of anti-apoptotic Bcl-2 protein and higher levels of pro-apoptotic Bax in the kidney with doses of 150 and 300 mg/kg in combination with oregano [94]. Remarkable decrease in hepatic enzymes including aspartate aminotransferase alanine (AST), aminotransferase (ALT), and alkaline phosphatase (ALP) with a parallel decrease in urea and creatinine were reported in diabetic rats treated with ethanol extract of chamomile flowers (500 mg/kg) [88]. Weight reduction was improved in alloxan-induced diabetic rats by administration of chamomile tea (1 g in 150 mL boiling water) for 8 weeks [93]. Extract of M. chamomilla and its compounds provided potent inhibitory activity against rat lens aldose reductase, advanced glycation end products, and DPPH radical. Additionally, isolated compounds including apigenin-7-O-β-Dglucoside, luteolin-7-O- β -D-glucoside, apigenin-7-O- β -D-glucuronide, luteolin-7-O-β-Dglucuronide, 3, 5-O-di-caffeoylquinic acid, apigenin, and luteolin suppressed sorbitol accumulation in rat lens in high glucose condition in vivo. These results indicated the beneficial effect of chamomile and its constituents in the prevention or treatment of diabetic complications [95].

Anti-obesity activity

Subchronic administration of decoction of chamomile flowers caused protection against high-fat diet obesity and an increase in hepatic, abdominal, and kidney fat weights in rats. The high-fat diet also caused an increase in the serum contents of TG, TC, and LDL and a decrease in HDL level which was significantly corrected by the administration of chamomile decoction [47].

Prevention of post-surgical adhesions

Postsurgical adhesion is a significant clinical problem after surgeries. Chamomile extracts reduced adhesion, fibrosis, inflammation, and post-surgical vascularization after surgery in female rats suggesting anti-inflammatory effects of the plant. The anti-inflammatory activity of chamomile resulted in the prevention of postsurgical adhesion [96].

Clinical Trials

Generalized anxiety disorder (GAD)

Participants with moderate to severe GAD were treated with pharmaceutical-grade chamomile extract (500 mg capsules 3 times a day) for 12 weeks. The responders were then randomized to either continuation of chamomile for an extra 26 weeks or a placebo. A greater number of placebo (25.5%) participants relapsed during follow-up versus chamomile (15.2%). Relapse means time for chamomile (11.4 weeks) was lower than relapse for placebo (6.3 weeks), and during follow-up, chamomile participants maintained significantly lower GAD symptoms than placebo with a significant reduction in body weight and blood arterial pressure [97]. Clinically meaningful and statistically significant improvement in GAD-7 score, anxiety, and wellbeing occurred in 11.7% of patients treated with chamomile extract (1500 mg per day for 8 weeks) [97]. In another study, patients with mild to moderate GAD were administered pharmaceutical grade German chamomile extract standardized to a content of 1.2% apigenin for 8 weeks and total Hamilton Anxiety Rating (HAM-A) scores reduced significantly versus placebo therapy. Positive changes in secondary outcomes included changes in the Beck Anxiety Inventory score, Psychological Well Being score, Clinical Global Impression Severity score, and the proportion of patients with \geq 50% reduction in baseline HAM-A score were also reported which suggests that chamomile has modest anxiolytic activity in patients with mild to moderate GAD [98]. In patients who had anxiety with depression, clinically meaningful changes over time in Hamilton Depression Rating (HAM-D) rating were observed indicating possible antidepressant activity of chamomile [99]. Further comparative studies of higher quality are needed to compare chamomile extract with conventional GAD therapies.

Efficacy in migraine

Traditional preparation containing 2% *M. chamomilla* essential oil rich in bisabolone oxide A (57.37%) and bisabolol oxide A (14.29%) was examined in patients suffering from migraine without aura. Pain based on visual analog scale (VAS) in the treatment group significantly decreased (beta= -0.38, p= 0.001). The nausea or vomiting decreased after 2 h of cutaneous application of the drug or placebo to 86.7% and 17.8%, respectively. Both photophobia- and phonophobia-associated attacks were decreased by chamomile preparation in patients [100].

Efficacy in attention-deficit disorder (ADHD)

The effect of *M. chamomilla* preparation containing 100 mg aqueous ethanol extract of the plant with 0.19 g levomenol (α -(-)-bisabolol) administered to the 3 patients suffering from ADHD was investigated for 4 weeks. Patients' mean scores for Conners' hyperactivity, inattention, and immaturity factors improved suggesting that chamomile may slightly be effective in ADHD [101].

Efficacy in oral health

One of the most common and disturbing side effects in high-dose chemotherapy is oral mucositis which causes erythema, edema, pain, and ulcers. A mouthwash containing M. chamomilla extract (1% w/v) with Mentha piperita oil (1% v/v) was administered to the patient undergoing chemotherapy and receiving a bone marrow transplant. The results showed that pain, dryness of the oral cavity, dysphagia, need for analgesics or total parental nutrition (TPN), duration of TPN, duration of oral mucositis, and maximum grade of oral mucositis significantly reduced in the intervention group in comparison with placebo [102]. In another study, chamomile mouthwash in patients who received 5flourouracil had no beneficial effect on the incidence, severity, and duration of oral mucositis [103]. A systematic review considered 11 clinical trials in which patients were treated mouthwash with chamomile when they underwent radiography and chemotherapy. Most of them showed a positive effect of M. chamomilla mouthwash in oral mucositis that may be caused through inhibition of nitric oxide and nitric oxide synthase, inhibition of COX-2 and MMP-9 with blocking the transcription factor NF-kß which reduce discomfort and

severity of oral mucositis [104].

The gingiva bleeding index was reduced by the administration of chamomile mouthwash in both gingivitis and chronic periodontitis [105]. In addition, plaque accumulation and gingival inflammation were lowered in patients who applied chamomile mouthwash [106]. In another mouthwash work, based on chamomile hydroalcoholic extract with chlorhexidine as control, both reduced bacterial plaque significantly, and chamomile had more plaque reduction effect which was not significantly different compared to the control [107]. More high-quality clinical trials to evaluate the positive effects of the mouthwash containing chamomile extract are warranted to indicate the duration of therapy to establish and maintain oral health.

Antidiabetic properties

Human studies reported changes in the levels of lipid profiles by administration of chamomile. Chamomile tea (3 g/150 mL hot water) three times per day for 8 weeks decreased the concentration of HbA1c, total cholesterol, triglyceride (TG), and low-density lipoprotein cholesterol (LDL-C) compared with the control group. In both groups, no significant changes were observed in serum high-density lipoprotein cholesterol (HDL-C) levels [108]. In contrast, no changes were observed in another study in total cholesterol, TG, HDL-C, and LDL-C by administration of three cups of chamomile tea to the patients who suffer from depression with type 2 diabetes. The level of HbA1c significantly reduced after the intervention [109].

Chamomile tea (3g/150 mL hot water) reduced malondialdehyde (MDA) levels and increased total antioxidant capacity, superoxide dismutase (SOD), catalase, and glutathione peroxidase (GPx) activities in patients who suffered from type 2 diabetes [110]. High-fat diet in rats increased renal and hepatic levels of MDA which was corrected by the administration of chamomile flower decoction [47]. The plant provided antiinflammatory activity in patients with diabetes by lowering tumor necrosis factor- α (TNF- α) and high sensitive-C reactive protein (hs-CRP) [111]. The decrease in serum creatinine was observed by intake of chamomile tea (10 g/100 mL boiling water) twice daily for 4 weeks in diabetic patients [112]. Results of a systematic review of 15 clinical studies showed novel functions of chamomile in the improvement of glycemic and

lipid profiles and oxidative stress indicators in diabetes mellitus and related complications [113].

Efficacy in menstrual problems

Chamomile capsules (100 mg) reduced the intensity of emotional symptoms more significantly than mefenamic acid after two cycle intervention in premenstrual syndrome (PMS). While intensity reduction of physical symptoms in both groups, chamomile and mefenamic acid, were not significantly different. The results of this study suggested that chamomile seems to be more effective than mefenamic acid in control of PMS-associated psychological pain [114]. Both chamomile and yarrow capsules reduced the severity of primary dysmenorrhea, while yarrow caused more reduction in pain severity [115].

The efficacy of chamomile in cyclic breast pain (mastalgia) was assessed for eight weeks. Chamomile preparation reduced mastalgia much more than placebo and was well tolerated and safe in the treatment of moderate mastalgia [116].

Efficacy in idiopathic hyperprolactinemia

In a randomized controlled clinical trial, women with idiopathic hyperprolactinemia was administered chamomile syrup (5 mL twice daily) or cabergoline tablet with a dose of 0.25 mg twice daily for 4 weeks. Results of the study indicated that cabergoline reduced mean prolactin levels significantly greater than that of chamomile [117].

Efficacy in enuresis

Chamomile has been used as a treatment of enuresis in traditional Persian medicine [118]. Aqueous extract of chamomile flowers was traditionally mixed with olive oil and boiled to evaporate the water portion. Chamomile oil which was prepared traditionally was applied on the perineal and suprapubic area of children at night. The mean frequency of enuresis was significantly reduced in the first, second, and third 2 weeks when compared with the base scores. No adverse effect was reported but some parents complained about the greasy nature of the herbal preparation [119]. The spasmolytic and anticholinergic activity of chamomile active constituents may be associated with the observed effect of traditional oil of the plant in children. Chamomile also showed spasmolytic activity in muscle tissues. Flavonoids can inhibit cAMP and cGMP phosphodiesterases which are considered as the mechanism of spasmolytic activity [77,82].

Conclusion

Matricaria chamomilla has been used since ancient times to treat different diseases. A wide range of secondary metabolites have been isolated and identified in the plant extracts and essential oil. Besides geographical features, the preparation of method of the essential oilinfluenced the composition and amount of the compounds [15,17,19]. Although the constituents of the plant oil were different in the previous studies, bisabolol oxide B, bisabolone oxide, and bisalolol oxide A, α -bisabolol oxide B, β farnesene, and chamazulene were the main components of the chamomile essential oil. Coumarins (like herniarin and umbelliferone), flavonoids (like quercetin and apigenin with their derivatives). polyacetylenes, and low concentrations of minerals such as calcium, potassium, magnesium, zinc, and cadmium were previously reported in chamomile aerial parts. Anti-inflammatory activity is one of the known properties of chamomile and its contributed mechanism was evaluated in vitro and in vivo. The anti-inflammatory activity of flavonoids, coumarins of the plant, and its volatile terpenoids like matricin and a-bisabolol mostly related to the reduction of TNF- α , IL-6, pro-inflammatory cytokines like IL-1 β , IL-8, suppressing the expression of COX-2, iNOS, 5-LOX, MMP-3, MMP-9, and p65 NF-κB with inhibiting the production of leukotriene B4 in vitro [120-125]. In animal models, these compounds reduced levels of TNF- α , NO, and IL-6, increased the MPO activity, reduced phosphorylation levels of ERK1/2, p38, and JNK, leukocyte migration, protein extravasations and TNF-α [122,124,126,127]. Chamomile preparation decreased oral mucositis, gingival inflammation, and increased antioxidant capacity with reduction of stress oxidative in type 2 diabetes in clinical [104,106,111,112]. studies Distinctive characteristic of chamomile, mechanisms of its biological activities and the constituents were evaluated and summarized in Tables 1-3 and discussed in the experimental for GAD, migraine, enuresis, menstrual problems, diabetes, mucositis, and ADHD. In vitro and experimental studies indicated antioxidant, anti-inflammation, antimicrobial. anti-diarrheal, gastroprotective, spasmolytic, hepatoprotective, anti-obesity, anti-platelet, antiangiogenic, cytotoxic,

prevention of post-surgical adhesions, and antidiabetic activity of the plant and its constituents. Although in vitro and in vivo studies deepen our knowledge of the mechanism of action of bioactive compounds, in vitro studies because of the lack of biokinetics in these systems and in vivo studies due to the differences in biokinetics parameters in lab animals, provide some drawbacks to extrapolating the obtained results to human [20,128]. The question is whether available data are enough to support the efficacy of the M. chamomilla preparations in clinical research. As mentioned, other studies rather than clinical trials can contribute to evaluating the mechanism of action of M. chamomilla extracts and its active components, while the final answer to the question about efficacy of the plant preparations in the humans' bodies as it was depicted in pre-clinical studies can only be clarified through well-designed clinical trials.

Acknowledgements

The authors gratefully thank Tehran University of Medical Sciences.

Author contributions

Azadeh Manayi and Mina Saeedi contributed to literature data collection and wrote the manuscript. Kasra Shahsavari contributed to drafting the manuscript. Mahnaz Khanavi supervised the study. All authors have read the final manuscript and approved the submission.

Declaration of interest

The authors declare that there is no conflict of interest. The authors alone are responsible for the accuracy and integrity of the paper content.

References

- Ruzicka J, Novak J. Mitochondrial genome variation between different accessions of *Matricaria chamomilla* L. (Asteraceae) based on SNP mutation analysis. *Genet Resour Crop Evol.* 2020; 67(4): 853–864.
- [2] Franke R, Schilcher H. Relevance and use of chamomile (*Matricaria recutita* L.). Acta *Hortic.* 2007; 749: 29–43.
- [3] Avonto C, Rua D, Lasonkar PB, Chittiboyina AG, Khan IA. Identification of a compound isolated from German chamomile (*Matricaria chamomilla*) with dermal sensitization potential. *Toxicol Appl*

Pharmacol. 2017; 318: 16-22.

- [4] Seyedjavadi SS, Khani S, Zare-Zardini H, Halabian R, Goudarzi M, Khatami S, Imani Fooladi AA, Amani J, Razzaghi-Abyaneh M. Isolation, functional characterization, and biological properties of MCh-AMP1, a novel antifungal peptide from *Matricaria chamomilla* L. *Chem Biol Drug Des*. 2019; 93(5): 949–959.
- [5] Zlabur JS, Zutić I, Radman S, Plesa M, Brnčić M, Barba FJ, Rocchetti G, Lucini L, Lorenzo JM, Domínguez R, Rimac Brnčić S, Galić A, Voća S. Effect of different green extraction methods and solvents on bioactive components of chamomile (*Matricaria chamomilla* L.) flowers. *Molecules*. 2020; 25(4): 1–17.
- [6] Singh O, Khanam Z, Misra N, Srivastava MK. Chamomile (*Matricaria chamomilla* L.): an overview. *Pharmacogn Rev.* 2011; 5(9): 82– 95.
- [7] Gupta V, Mittal P, Bansal P, Khokra SL, Kaushik D. Pharmacological potential of *Matricaria recutita*-a review. J Int J Pharm Sci Drug Res. 2010; 2(1): 12–16.
- [8] McKay DL, Blumberg JB. A review of the bioactivity and potential health benefits of chamomile tea (*Matricaria recutita* L.). *Phytother Res.* 2006; 20(7): 519–530.
- [9] Al-Snafi AE, Hasham LF. Bioactive constituents and pharmacological importance of *Matricaria chamomilla*: a recent review. *GSC Biol Pharm Sci.* 2023; 22(2): 79–98.
- [10] Ivanović S, Pajić M, Marković T. Economic effectiveness of mechanized harvesting of chamomile. *Econ Agric*. 2014; 61(2): 319– 330.
- [11] Mavandi P, Assareh MH, Dehshiri A, Rezadoost H, Abdossi V. Flower biomass, essential oil production and chemotype identification of some iranian *Matricaria chamomilla* var. *recutita* (L.) accessions and commercial varieties. J Essent Oil-Bear Plants. 2019; 22(5): 1228–1240.
- [12] Berechet MD, Manaila E, Stelescu MD, Craciun G. The composition of essential oils obtained from *Achillea millefolium* and *Matricaria chamomilla* L., originary from Romania. *Rev Chim.* 2017; 68(12): 2787– 2795.
- [13] Stanojevic LP, Marjanovic-Balaban ZR, Kalaba VD, Stanojevic JS, Cvetkovic DJ. Chemical composition, antioxidant and

antimicrobial activity of chamomile flowers essential oil (*Matricaria chamomilla* L.). J Essent Oil-Bear Plants. 2016; 19(8): 2017– 2028.

- [14] Rezaeih KAP, Gurbuz B, Uyanik M, Rahimi A, Arslan N. Volatile constituents variability in *Matricaria chamomilla* L. from Ankara, Turkey. J Essent Oil-Bear Plants. 2015; 18(1): 255–260.
- [15] Pirzad A, Alyari H, Shakiba M, Zehtab-Salmasi S, Mohammadi A. Essential oil content and composition of German chamomile (*Matricaria chamomilla* L.) at different irrigation regimes. J Agron. 2006; 5(3): 451–455.
- [16] Pino JA, Bayat F, Marbot R, Aguero J. Essential oil of chamomile *Chamomilla recutita* (L.) Rausch. from Iran. *J Essent Oil Res.* 2002; 14(6): 407–408.
- [17] Homami SS, Jaimand K, Rezaee MB, Afzalzadeh R. Comparative studies of different extraction methods of essential oil from *Matricaria recutita* L. in Iran. J Chil Chem Soc. 2016; 61(2): 2982–2984.
- [18] Baghalian K, Haghiry A, Naghavi MR, Mohammadi AJSH. Effect of saline irrigation water on agronomical and phytochemical characters of chamomile (*Matricaria recutita* L.). *Sci Hortic*. 2008; 116(4): 437–441.
- [19] Rahmati M, Azizi M, Khayyat MH, Nemati H, Asili J. Yield and oil constituents of chamomile (*Matricaria chamomilla* L.) flowers depending on nitrogen application, plant density and climate conditions. *J Essent Oil-Bear Plants*. 2011; 14(6): 731–741.
- [20] Manayi A, Saeidnia S. A concern on phthalate pollution of herbal extracts/medicines and detection methods. *Res J Pharmacogn*. 2015; 2(3): 49–54.
- [21] Manayi A, Kurepaz-Mahmoodabadi M, Gohari AR, Ajani Y, Saeidnia S. Presence of phthalate derivatives in the essential oils of a medicinal plant *Achillea tenuifolia*. *Daru*. 2014; 22(1): 1–6.
- [22] Omidpanah S, Saeidnia S, Saeedi M, Hadjiakhondi A, Manayi A. Phthalate contamination of some plants and herbal products. *Bol Latinoam Caribe Plantas Med Aromát*. 2018; 17(1): 61–67.
- [23] Avonto C, Wang M, Chittiboyina AG, Avula B, Zhao J, Khan IA. Hydroxylated bisabolol oxides: evidence for secondary oxidative metabolism in *Matricaria*

chamomilla. J Nat Prod. 2013; 76(10): 1848–1853.

- [24] Fonseca FN, Tavares MF, Horváth C. Capillary electrochromatography of selected phenolic compounds of *Chamomilla recutita*. *J Chromatogr A*. 2007; 1154(1-2): 390–399.
- [25] Weber B, Herrmann M, Hartmann B, Joppe H, Schmidt CO, Bertram HJ. HPLC/MS and HPLC/NMR as hyphenated techniques for accelerated characterization of the main constituents in chamomile (*Chamomilla recutita* [L.] Rauschert). Eur Food Res Technol. 2008; 226(4): 755–760.
- [26] Kanamori H, Terauchi M, Fuse JI, Sakamoto I. Studies on the evaluation of *Chamomillae* flos (part 1). Simultaneous and quantitative analysis of fat-soluble compounds. *Japanese J Pharmacogn*. 1992; 46(4): 384–388.
- [27] Mulinacci N, Romani A, Pinelli P, Vincieri F, Prucher D. Characterization of *Matricaria recutita* L. flower extracts by HPLC-MS and HPLC-DAD analysis. *Chromatographia*. 2000; 51(5-6): 301–307.
- [28] Zhao Y, Sun P, Ma Y, Wang K, Chang X, Bai Y, Zhang D, Yang L. Simultaneous quantitative determination of six caffeoylquinic acids in *Matricaria chamomilla* L. with high-performance liquid chromatography. J Chem. 2019; Article ID 4352832.
- [29] The Metabolomics Innovation Centre (TMIC). [Accessed 2012]. Available from: https://foodb.ca/compounds.
- [30] Kotov AG, Khvorost PP, Komissarenko NF. Coumarins of *Matricaria recutita*. *Chem Nat Compd*. 1992; 27(6): 753.
- [31] Redaelli C, Formentini L, Santaniello E. HPLC determination of coumarins in *Matricaria chamomilla*. *Planta Med.* 1981; 43(12): 412–413.
- [32] Molnar M, Mendešević N, Šubarić D, Banjari I, Jokić S. Comparison of various techniques for the extraction of umbelliferone and herniarin in *Matricaria chamomilla* processing fractions. *Chem Cent* J. 2017; 11: 1–8.
- [33] Repčák M, Imrich J, Franeková M. Umbelliferone, a stress metabolite of *Chamomilla recutita* (L.) Rauschert. *J Plant Physiol.* 2001; 158(8): 1085–1087.
- [34] Petrul'ová-Poracká V, Repčák M, Vilková M, Imrich J. Coumarins of *Matricaria chamomilla* L.: aglycones and glycosides.

Food Chem. 2013; 141(1): 54-59.

- [35] Haghi G, Hatami A, Safaei A, Mehran M. Analysis of phenolic compounds in *Matricaria chamomilla* and its extracts by UPLC-UV. *Res Pharm Sci.* 2014; 9(1): 31– 37.
- [36] Kunde R, Isaac O. On the flavones of chamomile (*Matricaria chamomilla* L.) and a new acetylated apigenin–7–glucoside. *Planta Med.* 1979; 37(10): 124–130.
- [37] Exner J, Reichling J, Cole T, Becker H. Methylated flavonoid aglycones from *"Matricariae* flos". *Planta Med.* 1981; 41(2): 198–200.
- [38] Carle R, Dölle B, Müller W, Baumeister U. Thermospray liquid chromatography-mass spectrometry (TSP LC-MS) analysis of acetylated apigenin 7-glucosides from *Chamomilla recutita. Planta Med.* 1992; 58: 686–687.
- [39] Redaelli C, Formentini L, Santaniello E. Apigenin 7-glucoside diacetates in ligulate flowers of *Matricaria chamomilla*. *Phytochemistry*. 1982; 21(7): 1828–1830.
- [40] Švehliková V, Bennett RN, Mellon FA, Needs PW, Piacente S, Kroon PA, Bao Y. Isolation, identification and stability of acylated derivatives of apigenin 7-Oglucoside from chamomile (*Chamomilla recutita* [L.] Rauschert). *Phytochemistry*. 2004; 65(16): 2323–2332.
- [41] Maday E, Szentmihályi K, Then M, Szőke É. Mineral element content of chamomile. *Acta Aliment*. 2000; 29(1): 51–57.
- [42] Chizzola R, Michitsch H, Mitteregger US. Extractability of selected mineral and trace elements in infusions of chamomile. *Int J Food Sci Nutr.* 2008; 59(6): 451–456.
- [43] Roby MHH, Sarhan MA, Selim KAH, Khalel KI. Antioxidant and antimicrobial activities of essential oil and extracts of fennel (*Foeniculum vulgare* L.) and chamomile (*Matricaria chamomilla* L.). Ind Crops Prod. 2013; 44: 437–445.
- [44] Al-Dabbagh B, Elhaty IA, Elhaw M, Murali C, Al Mansoori A, Awad B, Amin A. Antioxidant and anticancer activities of chamomile (*Matricaria recutita* L.). *BMC Res Notes*. 2019; 12(1): 1–8.
- [45] Hajaji S, Alimi D, Jabri MA, Abuseir S, Gharbi M, Akkari H. Anthelmintic activity of *Tunisian chamomile (Matricaria recutita L.)* against *Haemonchus contortus*. J Helminthol.

2018; 92(2): 168–177.

- [46] Sebai H, Jabri MA, Souli A, Hosni K, Rtibi K, Tebourbi O, El-Benna J, Sakly M. Chemical composition, antioxidant properties and hepatoprotective effects of chamomile (*Matricaria recutita* L.) decoction extract against alcohol-induced oxidative stress in rat. *Gen Physiol Biophys.* 2015; 34(3): 263–275.
- [47] Jabri MA, Sakly M, Marzouki L, Sebai H. Chamomile (*Matricaria recutita* L.) decoction extract inhibits in vitro intestinal glucose absorption and attenuates high fat diet-induced lipotoxicity and oxidative stress. *Biomed Pharmacother*. 2017; 87: 153–159.
- [48] Kazemi M. Chemical composition and antimicrobial activity of essential oil of *Matricaria recutita*. Int J Food Prop. 2015; 18(8): 1784–1792.
- [49] Pauli A. α-Bisabolol from chamomile–A specific ergosterol biosynthesis inhibitor? *Int J Aromather*. 2006; 16(1): 21–25.
- [50] Cvetanović A, Švarc-Gajić J, Zeković Z, Savić S, Vulić J, Mašković P, Ćetković G. Comparative analysis of antioxidant, antimicrobiological and cytotoxic activities of native and fermented chamomile ligulate flower extracts. *Planta*. 2015; 242(3): 721– 732.
- [51] Marino M, Bersani C, Comi G. Impedance measurements to study the antimicrobial activity of essential oils from Lamiaceae and Compositae. *Int J Food Microbiol.* 2001; 67(3): 187–195.
- [52] Hyldgaard M, Mygind T, Meyer RL. Essential oils in food preservation: mode of action, synergies, and interactions with food matrix components. *Front Microbiol*. 2012; 3: 1–24.
- [53] Calo JR, Crandall PG, O'Bryan CA, Ricke SC. Essential oils as antimicrobials in food systems–a review. *Food Control.* 2015; 54: 111–119.
- [54] Mekonnen A, Yitayew B, Tesema A, Taddese S. In vitro antimicrobial activity of essential oil of Thymus schimperi, Matricaria chamomilla, Eucalyptus globulus, and Rosmarinus officinalis. Int J Microbiol. 2016; Article ID 9545693.
- [55] Nogueira JCR, Diniz MFM, Lima EO. In vitro antimicrobial activity of plants in acute otitis externa. Braz J Otorhinolaryngol. 2008; 74(1): 118–124.
- [56] Fabri R, Nogueira M, Dutra L, Bouzada M,

Scio E. Antioxidant and antimicrobial potential of Asteraceae species. *Rev Bras de Plantas Medicinais*. 2011; 13(2): 183–189.

- [57] Bensch K, Tiralongo J, Schmidt K, Matthias A, Bone K, Lehmann R, Tiralongo E. Investigations into the antiadhesive activity of herbal extracts against *Campylobacter jejuni*. *Phytother Res.* 2011; 25(8): 1125–1132.
- [58] Cwikla C, Schmidt K, Matthias A, Bone K, Lehmann R, Tiralongo E. Investigations into the antibacterial activities of phytotherapeutics against *Helicobacter pylori* and *Campylobacter jejuni*. *Phytother Res*. 2010; 24(5): 649–656.
- [59] Móricz AM, Szarka S, Ott PG, Héthelyi EB, Szoke E, Tyihák E. Separation and identification of antibacterial chamomile components using OPLC, bioautography and GC-MS. *Med Chem.* 2012; 8(1): 85–94.
- [60] Jesionek W, Móricz ÁM, Ott PG, Kocsis B, Horváth G, Choma IM. TLC-direct bioautography and LC/MS as complementary methods in identification of antibacterial agents in plant tinctures from the Asteraceae family. J AOAC Int. 2015; 98(4): 857–861.
- [61] Hajaji S, Sifaoui I, López-Arencibia A, Reyes-Batlle M, Jiménez IA, Bazzocchi IL, Valladares B, Akkari H, Lorenzo-Morales J, Piñero JE. Leishmanicidal activity of αbisabolol from Tunisian chamomile essential oil. *Parasitol Res.* 2018; 117(9): 2855–2867.
- [62] Morales-Yuste M, Morillas-Márquez F, Martín-Sánchez J, Valero-López A, Navarro-Moll MC. Activity of (-)α-bisabolol against *Leishmania infantum* promastigotes. *Phytomedicine*. 2010; 17(3): 279–281.
- [63] Baldissera MD, Grando TH, de Souza CF, Cossetin LF, da Silva APT, Giongo JL, Monteiro SG. A nanotechnology based new approach for *Trypanosoma evansi* chemotherapy: *In vitro* and *vivo* trypanocidal effect of (-)-α-bisabolol. *Exp Parasitol*. 2016; 170: 156–160.
- [64] Hajaji S, Sifaoui I, López-Arencibia A, Reyes-Batlle M, Jiménez IA, Bazzocchi IL, Valladares B, Pinero JE, Lorenzo-Morales J, Akkari H. Correlation of radical-scavenging capacity and amoebicidal activity of *Matricaria recutita* L. (Asteraceae). *Exp Parasitol*. 2017; 183: 212–217.
- [65] Caleja C, Ribeiro A, Barros L, Barreira JC, Antonio AL, Oliveira MBPP, Barreiro MF,

Ferreira IC. Cottage cheeses functionalized with fennel and chamomile extracts: comparative performance between free and microencapsulated forms. *Food Chem.* 2016; 199: 720–726.

- [66] Caleja C, Barros L, Antonio AL, Ciric A, Barreira JC, Sokovic M, Oliveira MBP, Santos-Buelga C, Ferreira Isabel CFR. Development of a functional dairy food: exploring bioactive and preservation effects of chamomile (*Matricaria recutita* L.). J Funct Foods. 2015; 16: 114–124.
- [67] Caleja C, Barros L, Antonio AL, Oliveira MBP, Ferreira IC. A comparative study between natural and synthetic antioxidants: evaluation of their performance after incorporation into biscuits. *Food Chem.* 2017; 216: 342–346.
- [68] Aliheidari N, Fazaeli M, Ahmadi R, Ghasemlou M, Emam-Djomeh Z. Comparative evaluation on fatty acid and *Matricaria recutita* essential oil incorporated into casein-based film. *Int J Biol Macromol.* 2013; 56: 69–75.
- [69] Soković M, Glamočlija J, Marin PD, Brkić D, Van Griensven LJ. Antibacterial effects of the essential oils of commonly consumed medicinal herbs using an *in vitro* model. *Molecules*. 2010; 15(11): 7532–7546.
- [70] Queiroz MBR, Lucena G, Caldas ED, Silva M. Evaluation of the anti-inflammatory activity of gel with *Matricaria recutita* L. using a permeation enhancer. *Rev Bras Farm*. 2014; 95(2): 676–694.
- [71] Srivastava JK, Pandey M, Gupta S. Chamomile, a novel and selective COX-2 inhibitor with anti-inflammatory activity. *Life Sci*. 2009; 85(19-20): 663–669.
- [72] Bijak M, Saluk J, Tsirigotis-Maniecka M, Komorowska H, Wachowicz B, Zaczyńska E, Czarny A, Czechowski F, Nowak P, Pawlaczyk I. The influence of conjugates isolated from *Matricaria chamomilla* L. on platelets activity and cytotoxicity. *Int J Biol Macromol.* 2013; 61: 218–229.
- [73] Danaei N, Kokhdan EP, Manzouri L, Nikseresht M. The effect of bevacizumab and hydroalcohlic extract of *Matricaria chamomilla* on cell viability and nitric oxide (NO) production in HT-29; a human colorectal cancer cell line. *Armaghane Danesh*. 2016; 20(12): 1107–1118.
- [74] Jabri MA, Aissani N, Tounsi H, Sakly M,

Marzouki L, Sebai H. Protective effect of chamomile (*Matricaria recutita* L.) decoction extract against alcohol-induced injury in rat gastric mucosa. *Pathophysiology*. 2017; 24(1): 1–8.

- [75] Sebai H, Jabri MA, Souli A, Rtibi K, Selmi S, Tebourbi O, El-Benna J, Sakly M. Antidiarrheal and antioxidant activities of chamomile (*Matricaria recutita* L.) decoction extract in rats. *J Ethnopharmacol.* 2014; 152(2): 327–332.
- [76] Yazdi H, Seifi A, Changizi S, Khori V, Hossini F, Davarian A, Jand Y, Enayati A, Mazandarani M, Nanvabashi F. Hydroalcoholic extract of *Matricaria recutita* exhibited dual anti-spasmodic effect via modulation of Ca(2+) channels, NO and PKA(2)-kinase pathway in rabbit jejunum. *Avicenna J Phytomed*. 2017; 7(4): 334–344.
- [77] Achterrath-Tuckermann U. Kunde R. О, Thiemer K. Flaskamp E, Isaac Pharmacological investigations with compounds of chamomile. V. Investigations on the spasmolytic effect of compounds of chamomile and Kamillosan on the isolated guinea pig ileum. Planta Med. 1980; 39(1): 38–50.
- [78] Forster H, Niklas H, Lutz S. Antispasmodic effects of some medicinal plants. *Planta Med*. 1980; 40(12): 309–319.
- [79] Rotondo A, Serio R, Mulè F. Gastric relaxation induced by apigenin and quercetin: analysis of the mechanism of action. *Life Sci.* 2009; 85(1-2): 85–90.
- [80] Hagelauer D, Kelber O, Weiser D, Heinle H. Effects of STW 5 (Iberogast®) on prostaglandinF2α–induced contractions of ileum of mice in-vitro. *Planta Med.* 2006; 72(11): 280.
- [81] Murthy KS. Signaling for contraction and relaxation in smooth muscle of the gut. *Annu Rev Physiol*. 2006; 68: 345–374.
- [82] Maschi O, Cero ED, Galli GV, Caruso D, Bosisio E, Dell'Agli M. Inhibition of human cAMP-phosphodiesterase as a mechanism of the spasmolytic effect of *Matricaria recutita* L. *J Agric Food Chem.* 2008; 56(13): 5015– 5020.
- [83] Amira S, Rotondo A, Mulè F. Relaxant effects of flavonoids on the mouse isolated stomach: structure-activity relationships. *Eur J Pharmacol.* 2008; 599(1-3): 126–130.
- [84] Gharzouli K, Holzer P. Inhibition of guinea

pig intestinal peristalsis by the flavonoids quercetin, naringenin, apigenin and genistein. *Pharmacology*. 2004; 70(1): 5–14.

- [85] Guimarães MV, Melo IM, Adriano Araújo VM, Tenazoa Wong DV, Roriz Fonteles CS, Moreira Leal LK, Ribeiro RA, Lima V. Dry extract of *Matricaria recutita* L. (Chamomile) prevents ligature-induced alveolar bone resorption in rats via inhibition of tumor necrosis factor-α and interleukin-1β. J Periodontol. 2016; 87(6): 706–715.
- [86] Prasanna R, Ashraf EA, Essam MA. Chamomile and oregano extracts synergistically exhibit antihyperglycemic, antihyperlipidemic, and renal protective effects in alloxan-induced diabetic rats. *Can J Physiol Pharmacol.* 2017; 95(1): 84–92.
- [87] Alouie A, Zehsaz F, Pouzesh Jadidi R. Effect of endurance exercise with *Chamomila recutita* leaves extract on liver superoxide dismutase activity and malondialdehyde levels in type 1 diabetic rats. *Res Med.* 2017; 40(4): 165–171.
- [88] Al-Musa H, Al-Hashem F. Hypoglycemic, hepato-renal and antioxidant potential effects of *Chamomila recutita* flowers ethanolic extract in streptozotocin-diabetic rats. *Am J Pharmacol Toxicol*. 2014; 9(1): 1–12.
- [89] Abdolmaleki F, Heidarianpour A. The response of serum glypican-4 levels and its potential regulatory mechanism to endurance training and chamomile flowers' hydroethanolic extract in streptozotocin–nicotinamide-induced diabetic rats. *Acta Diabetol.* 2018; 55(9): 935–942.
- [90] Kato A, Minoshima Y, Yamamoto J, Adachi I, Watson AA, Nash RJ. Protective effects of dietary chamomile tea on diabetic complications. *J Agric Food Chem.* 2008; 56(17): 8206–8211.
- [91] Cemek M, Kağa S. Şimşek N. Büyükokuroğlu ME, Konuk M. Antihyperglycemic and antioxidative potential of Matricaria chamomilla L. in streptozotocin-induced diabetic rats. J Nat Med. 2008; 62(3): 284-293.
- [92] Eddouks M, Lemhadri A, Zeggwagh N, Michel J. Potent hypoglycaemic activity of the aqueous extract of *Chamaemelum nobile* in normal and streptozotocin-induced diabetic rats. *Diabetes Res Clin Pract*. 2005; 67(3): 189–195.
- [93] Khan SS, Najam R, Anser H, Riaz B, Alam

N. Chamomile tea: herbal hypoglycemic alternative for conventional medicine. *Pak J Pharm Sci.* 2014; 27(5): 1509–1514.

- [94] Prasanna R, Ashraf EA, Essam MA. Chamomile and oregano extracts synergistically exhibit antihyperglycemic, antihyperlipidemic, and renal protective effects in alloxan-induced diabetic rats. *Can J Physiol Pharmacol*. 2017; 95(1): 84–92.
- [95] Hwang SH, Wang Z, Guillen Quispe YN, Lim SS. Evaluation of aldose reductase, protein glycation, and antioxidant inhibitory activities of bioactive flavonoids in *Matricaria recutita* L. and their structureactivity relationship. J Diabetes Res. 2018; Article ID 3276162.
- [96] Rajaei M, Asadi I. Fibrinolytic effects of Matricaria chamomila in preventing peritoneal adhesions. Bull Env Pharmacol Life Sci. 2014; 3(5): 40–45.
- [97] Mao JJ, Xie SX, Keefe JR, Soeller I, Li QS, Amsterdam JD. Long-term chamomile (*Matricaria chamomilla* L.) treatment for generalized anxiety disorder: a randomized clinical trial. *Phytomedicine*. 2016; 23(14): 1735–1742.
- [98] Amsterdam JD, Li Y, Soeller I, Rockwell K, Mao JJ, Shults J. A randomized, double-blind, placebo-controlled trial of oral *Matricaria recutita* (chamomile) extract therapy for generalized anxiety disorder. *Sci Rep.* 2009; 29(4): 378–382.
- [99] Amsterdam JD, Shults J, Soeller I, Mao JJ, Rockwell K, Newberg AB. Chamomile (*Matricaria recutita*) may provide antidepressant activity in anxious, depressed humans: an exploratory study. *Alt Ther Health Med.* 2012; 18(5): 44–49.
- [100] Zargaran A, Borhani-Haghighi A, Salehi-Marzijarani M, Faridi P, Daneshamouz S, Azadi A, Sadeghpour H, Sakhteman A, Mohagheghzadeh A. Evaluation of the effect of topical chamomile (*Matricaria chamomilla* L.) oleogel as pain relief in migraine without aura: a randomized, doubleblind, placebo-controlled, crossover study. *Neurol Sci.* 2018; 39(8): 1345–1353.
- [101] Niederhofer H. Observational study: Matricaria chamomilla may improve some symptoms of attention-deficit hyperactivity disorder. Phytomedicine. 2009; 16(4): 284– 286.
- [102] Tavakoli Ardakani M, Ghassemi S,

Mehdizadeh M, Mojab F, Salamzadeh J, Ghassemi S, Hajifathali A. Evaluating the effect of *Matricaria recutita* and *Mentha piperita* herbal mouthwash on management of oral mucositis in patients undergoing hematopoietic stem cell transplantation: a randomized, double blind, placebo controlled clinical trial. *Complement Ther Med.* 2016; 29: 29–34.

- [103] Fidler P, Loprinzi CL, O'Fallon JR, Leitch JM, Lee JK, Hayes DL, Novotny P, Clemens-Schutjer D, Bartel J, Michalak JC. Prospective evaluation of a chamomile mouthwash for prevention of 5-FU-induced oral mucositis. *Cancer.* 1996; 77(3): 522–525.
- [104] Gomes VTS, Nonato Silva Gomes R, Gomes MS, Joaquim WM, Lago EC, Nicolau RA. Effects of *Matricaria recutita* (L.) in the treatment of oral mucositis. *Sci World J*. 2018; Article ID 4392184.
- [105] Batista ALA, Lins RDAU, de Souza Coelho R, do Nascimento Barbosa D, Belém NM, Celestino FJA. Clinical efficacy analysis of the mouth rinsing with pomegranate and chamomile plant extracts in the gingival bleeding reduction. *Complement Ther Clin Pract*. 2014; 20(1): 93–98.
- [106] Pourabbas R, Delazar A. The effect of German chamomile mouthwash on dental plaque and gingival inflammation. *Iran J Pharm Res.* 2010; 4(2): 105–109.
- [107] Lins R, Vasconcelos F, Leite R, Coelho-Soares R, Barbosa D. Clinical evaluation of mouthwash with extracts from aroeira (*Schinus terebinthifolius*) and chamomile (*Matricaria recutita* L.) on plaque and gingivitis. *Rev Bras de Plantas Medicinais*. 2013; 15(1): 112–120.
- [108] Rafraf M, Zemestani M, Asghari-Jafarabadi M. Effectiveness of chamomile tea on glycemic control and serum lipid profile in patients with type 2 diabetes. *J Endocrinol Invest.* 2015; 38(2): 163–170.
- [109] Kermanian S, Mozaffari-Khosravi H, Dastgerdi G, Zavar-Reza J, Rahmanian M. The effect of chamomile tea versus black tea on glycemic control and blood lipid profiles in depressed patients with type 2 diabetes: a randomized clinical trial. *J Nutr Foof Secur*. 2018; 3(3): 157–166.
- [110] Zemestani M, Rafraf M, Asghari-Jafarabadi M. Chamomile tea improves glycemic indices and antioxidants status in

patients with type 2 diabetes mellitus. *Nutrition*. 2016; 32(1): 66–72.

- [111] Zemestani M, Rafraf M, Asghari-Jafarabadi M. Effects of chamomile tea on inflammatory markers and insulin resistance in patients with type 2 diabetes mellitus. *Trends Gen Pract*. 2018; 1(3): 1–6.
- [112] Kaseb F, Yazdanpanah Z, Biregani AN, Yazdi NB, Yazdanpanah Z. The effect of chamomile (*Matricaria recutita* L.) infusion on blood glucose, lipid profile and kidney function in type 2 diabetic patients: a randomized clinical trial. *Prog Food Nutr Sci.* 2018; 20(S1): 110–118.
- [113] Hajizadeh-Sharafabad F, Varshosaz P, Jafari-Vayghan H, Alizadeh M, Maleki V. Chamomile (*Matricaria recutita* L.) and diabetes mellitus, current knowledge and the way forward: a systematic review. *Complement Ther Med.* 2020; Article ID 102284.
- [114] Sharifi F, Simbar M, Mojab F, Majd HA. Comparison of the effects of *Matricaria chamomila* (chamomile) extract and mefenamic acid on the intensity of premenstrual syndrome. *Complement Ther Clin Pract*. 2014; 20(1): 81–88.
- [115] Radfar S, Shahoie R, Noori B, Jalilian F, Nasab LH. Comparative study on the effect of *Matricaria chamomile* and *Achillea millefolium* capsules on primary dysmenorrhea intensity of dormitory students of kurdistan university of medical sciences, 2018. J Pharm Res Int. 2018; 25(3): 1–7.
- [116] Saghafi N, Rhkhshandeh H, Pourmoghadam N, Pourali L, Ghazanfarpour M, Behrooznia A, Vafisani F. Effectiveness of *Matricaria chamomilla* (chamomile) extract on pain control of cyclic mastalgia: a double-blind randomised controlled trial. J Obstetr Gynaecol. 2018; 38(1): 81–84.
- [117] Kabiri M, Kamalinejad M, Bioos S, Shariat M, Sohrabvand F. Comparative study of the effects of chamomile (*Matricaria chamomilla* L.) and cabergoline on idiopathic hyperprolactinemia: a pilot randomized controlled trial. *Iran J Pharm Res.* 2019; 18(3): 1612–1621.
- [118] Aghili Khorasani M. Makhzan al advieh. Tehran: Bavardaran Press, 2001.
- [119] Sharifi H, Minaie MB, Qasemzadeh MJ, Ataei N, Gharehbeglou M, Heydari M. Topical use of *Matricaria recutita* L.

(chamomile) oil in the treatment of monosymptomatic enuresis in children: a double-blind randomized controlled trial. *Evid Based Complement Alternat Med.* 2017; 22(1): 12–17.

- [120] Comalada M, Ballester I, Bailon E, Sierra S, Xaus J, Gálvez J, de Medina FS, Zarzuelo A. Inhibition of pro-inflammatory markers in primary bone marrow-derived mouse macrophages by naturally occurring flavonoids: analysis of the structure–activity relationship. *Biochem Pharmacol.* 2006; 72(8): 1010–1021.
- [121] Liang YC, Huang YT, Tsai SH, Lin-Shiau SY, Chen CF, Lin JK. Suppression of inducible cyclooxygenase and inducible nitric oxide synthase by apigenin and related flavonoids in mouse macrophages. *Carcinogenesis*. 1999; 20(10): 1945–1952.
- [122] Park JS, Kim DK, Shin HD, Lee HJ, Jo HS, Jeong JH, Choi YL, Lee CJ, Hwang SC. Apigenin regulates interleukin-1β-induced production of matrix metalloproteinase both in the knee joint of rat and in primary cultured articular chondrocytes. *Biomol Ther*. 2016; 24(2): 163–170.
- [123] Flemming M, Kraus B, Rascle A, Jürgenliemk G, Fuchs S, Fürst R, Heilmann J. Revisited anti-inflammatory activity of matricine *in vitro*: comparison with chamazulene. *Fitoterapia*. 2015; 106: 122– 128.
- [124] Salehcheh M, Safari O, Khodayar MJ, Mojiri-Forushani H, Cheki M. The protective effect of herniarin on genotoxicity and apoptosis induced by cisplatin in bone marrow cells of rats. *Drug Chem Toxicol*. 2022; 45(4): 1470–1475.
- [125] Safayhi H, Sabieraj J, Sailer ER, Ammon HPT. Chamazulene: an antioxidant-type inhibitor of leukotriene B4 formation. *Planta Med.* 1994; 60(5): 410–413.
- [126] Chang X, He H, Zhu L, Gao J, Wei T, Ma Z, Yan T. Protective effect of apigenin on Freund's complete adjuvant-induced arthritis in rats via inhibiting P2X7/NF-κB pathway. *Chem Biol Interact*. 2015; 236: 41–46.
- [127] Vasconcelos JF, Teixeira MM, Barbosa-Filho JM, Agra MF, Nunes XP, Giulietti AM, Ribeiro-Dos-Santos R, Soares MB. Effects of umbelliferone in a murine model of allergic airway inflammation. *Eur J Pharmacol.* 2009; 609(1): 126–131.

- [128] Sánchez M, González-Burgos E, Gómez-Serranillos MP. The pharmacology and clinical efficacy of *Matricaria recutita* L.: a systematic review of *in vitro*, *in vivo* studies and clinical trials. *Food Rev Int*. 2022; 10(8): 1668–1702.
- [129] Cavalcante HAO, Silva-Filho SE, Wiirzler LAM, Cardia GFE, Uchida NS, Silva-Comar FMS, Bersani-Amado CA, Cuman RKN. Effect of (-)-α-bisabolol on the inflammatory response in systemic infection experimental model in C57BL/6 mice. *Inflammation*. 2020; 43(1): 193–203.
- [130] Xu C, Sheng S, Dou H, Chen J, Zhou K, Lin Y, Yang H. α-Bisabolol suppresses the inflammatory response and ECM catabolism in advanced glycation end products-treated chondrocytes and attenuates murine osteoarthritis. *Int Immunopharmacol.* 2020; Article ID 106530.
- [131] D'Almeida APL, Pacheco de Oliveira MT, de Souza ÉT, de Sá Coutinho D, Ciambarella BT, Gomes CR, Terroso T, Guterres SS, Pohlmann AR, Silva PM, Martins MA, Bernardi A. α-Bisabolol-loaded lipid-core nanocapsules reduce lipopolysaccharideinduced pulmonary inflammation in mice. *Int J Nanomedicine*. 2017; 12: 4479–4491.
- [132] Rocha NF, Rios ER, Carvalho AM, Cerqueira GS, Lopes Ade A, Leal LK, Dias ML, de Sousa DP, de Sousa FC. Antinociceptive and anti-inflammatory activities of (-)-α-bisabolol in rodents. *Naunyn Schmiedebergs Arch Pharmacol.* 2011; 384(6): 525–533.
- [133] Pereira da Cruz R, Sampaio de Freitas T, Socorro Costa MD, Lucas Dos Santos AT, Ferreira Campina F, Pereira RLS, Bezerra JWA, Quintans-Júnior LJ, De Souza Araújo AA, Júnior JPS, Iriti M, Varoni EM, Menezes IRA, Melo Coutinho HD, Morais-Braga MFB. Effect of α -bisabolol and its β cyclodextrin complex as tetk and nora efflux pump inhibitors in *Staphylococcus aureus* strains. *Antibiotics*. 2020; 9(1): 1–8.
- [134] Hajaji S, Sifaoui I, López-Arencibia A, Reyes-Batlle M, Jiménez IA, Bazzocchi IL, Valladares B, Akkari H, Lorenzo-Morales J, Piñero JE. Leishmanicidal activity of αbisabolol from *Tunisian chamomile* essential oil. *Parasitol Res.* 2018; 117(9): 2855–2867.
- [135] Anter J, Romero-Jiménez M, Fernández-Bedmar Z, Villatoro-Pulido M, Analla M,

Alonso-Moraga A, Muñoz-Serrano A. Antigenotoxicity, cytotoxicity, and apoptosis induction by apigenin, bisabolol, and protocatechuic acid. *J Med Food*. 2011; 14(3): 276–283.

- [136] Aron de Miranda HA, Gonçalves JCR, Cruz JS, Araújo DAM. Evaluation of the sesquiterpene (-)-α-bisabolol as a novel peripheral nervous blocker. *Neurosci Lett.* 2010; 472(1): 11–15.
- [137] El-Lakany SA, Abd-Elhamid AI, Kamoun EA, El-Fakharany EM, Samy WM, Elgindy NA. α-Bisabolol-loaded cross-linked zein nanofibrous 3d-scaffolds for accelerating wound healing and tissue regeneration in rats. *Int J Nanomedicine*. 2019; 14: 8251–8270.
- [138] Fernandes MYD, Carmo MRSD, Fonteles AA, Neves JCS, Silva ATAD, Pereira JF, Ferreira EO, Lima NMR, Neves KRT, Andrade GM. (-)-α-Bisabolol prevents neuronal damage and memory deficits through reduction of proinflammatory markers induced by permanent focal cerebral ischemia in mice. Eur J Pharmacol. 2019; 842: 270-280.
- [139] Sampaio TL, Menezes RR, da Costa MF, Meneses GC, Arrieta MC, Chaves Filho AJ, de Morais GB, Libório AB, Alves RS, Evangelista JS, Martins AM. Nephroprotective effects of (-)-α-bisabolol against ischemic-reperfusion acute kidney injury. *Phytomedicine*. 2016; 23(14): 1843– 1852.
- [140] Fukunaga E, Hirao Y, Ogata-Ikeda I, Nishimura Y, Seo H, Oyama Y. Bisabololoxide A, one of the constituents in german chamomile extract, attenuates cell death induced by calcium overload. *Phytother Res.* 2014; 28(5): 685–691.
- [141] Ogata-Ikeda I, Seo H, Kawanai T, Hashimoto E, Oyama Y. Cytotoxic action of bisabololoxide A of German chamomile on human leukemia K562 cells in combination with 5-fluorouracil. *Phytomedicine*. 2011; 18(5): 362–365.
- [142] Ogata I, Kawanai T, Hashimoto E, Nishimura Y, Oyama Y, Seo H. Bisabololoxide A, one of the main constituents in German chamomile extract, induces apoptosis in rat thymocytes. *Arch Toxicol.* 2010; 84(1): 45–52.
- [143] Kobayashi Y, Suzuki A, Kobayashi A, Kasai A, Ogata Y, Kumada Y. Suppression

of sensory irritation by chamomile essential oil and its active component-bisabololoxide A. *Acta Hortic*. 2007; 749: 163–174.

- [144] Fang K, Wang L, Chen L, Liu T, Fang Z. Antiproliferative effects of matricine in gemcitabine-resistant human pancreatic carcinoma cells are mediated via mitochondrial-mediated apoptosis, inhibition of cell migration, invasion suppression, and mammalian target of rapamycin (mTOR)-TOR/PI3K/AKT signalling pathway. *Med Sci Monit*. 2019; 25: 2943–2949.
- [145] Capuzzo A, Occhipinti A, Maffei ME. Antioxidant and radical scavenging activities of chamazulene. *Nat Prod Res.* 2014; 28(24): 2321–2323.
- [146] Querio G, Antoniotti S, Foglietta F, Bertea CM, Canaparo R, Gallo MP, Levi R. Chamazulene attenuates ROS levels in bovine aortic endothelial cells exposed to high glucose concentrations and hydrogen peroxide. *Front Physiol.* 2018; 9: 1–7.
- [147] Funakoshi-Tago M, Nakamura K, Tago K, Mashino T, Kasahara T. Anti-inflammatory activity of structurally related flavonoids, apigenin, luteolin and fisetin. *Int Immunopharmacol.* 2011; 11(9): 1150–1159.
- [148] Kim HK, Cheon BS, Kim YH, Kim SY, Kim HP. Effects of naturally occurring flavonoids on nitric oxide production in the macrophage cell line RAW 264.7 and their structure–activity relationships. *Biochem Pharmacol.* 1999; 58(5): 759–765.
- [149] Nicholas C, Batra S, Vargo MA, Voss OH, Gavrilin MA, Wewers MD, Guttridge DC, Grotewold E, Doseff AI. Apigenin blocks lipopolysaccharide-induced lethality in vivo and proinflammatory cytokines expression by inactivating NF-κB through the suppression of p65 phosphorylation. *J Immunol.* 2007; 179(10): 7121–7127.
- [150] Soliman KF, Mazzio EA. In vitro attenuation of nitric oxide production in C6 astrocyte cell culture by various dietary compounds. Proc Soc Exp Biol Med. 1998; 218(4): 390–397.
- [151] Rudan I, Sidhu S, Papana A, Meng SJ, Xin-Wei Y, Wang W, Campbell-Page RM, Demaio AR, Nair H, Sridhar D, Theodoratou E, Dowman B, Adeloye D, Majeed A, Car J, Campbell H, Wang W, Chan KY. Prevalence of rheumatoid arthritis in low-and middle– income countries: a systematic review and

analysis. J Glob Health. 2015; 5(1): 1–10.

- [152] Romanova D, Vachalkova A, Cipak L, Ovesna Z, Rauko P. Study of antioxidant effect of apigenin, luteolin and quercetin by DNA protective method. *Neoplasma*. 2001; 48(2): 104–107.
- [153] Kuo ML, Lee KC, Lin JK. Genotoxicities of nitropyrenes and their modulation by apigenin, tannic acid, ellagic acid and indole-3-carbinol in the Salmonella and CHO systems. *Mutat Res.* 1992; 270(2): 87–95.
- [154] Van Dross R, Xue Y, Knudson A, Pelling JC. The chemopreventive bioflavonoid apigenin modulates signal transduction pathways in keratinocyte and colon carcinoma cell lines. J Nutr. 2003; 133(11 Suppl 1): 3800S–3804S.
- [155] Birt DF, Mitchell DL, Gold B, Pour P, Pinch HC. Inhibition of ultraviolet light induced skill carcinogenesis in SKH-1 mice by apigenin, a plant flavonoid. *Anticancer Res.* 1997; 17(1A): 85–91.
- [156] Dong J, Qiu J, Wang J, Li H, Dai X, Zhang Y, Wang X, Tan W, Niu X, Deng X, Zhao S. Apigenin alleviates the symptoms of *Staphylococcus aureus* pneumonia by inhibiting the production of alpha-hemolysin. *FEMS Microbiol Lett.* 2013; 338(2): 124–131.
- [157] Xia F, Li X, Wang B, Gong P, Xiao F, Yang M, Zhang L, Song J, Hu L, Cheng M, Sun C, Feng X, Lei L, Ouyang S, Liu ZJ, Li X, Gu J, Han W. Combination therapy of LysGH15 and apigenin as a new strategy for treating pneumonia caused by *Staphylococcus aureus*. *Appl Environ Microbiol*. 2016; 82(1): 87–94.
- [158] Kuo CH, Weng BC, Wu CC, Yang SF, Wu DC, Wang YC. Apigenin has anti-atrophic gastritis and anti-gastric cancer progression effects in Helicobacter pylori-infected Mongolian gerbils. *J Ethnopharmacol.* 2014; 151(3): 1031–1039.
- [159] Koo H, Schobel B, Scott-Anne K, Watson G, Bowen W, Cury J, Rosalen PL, Park YK. Apigenin and tt-farnesol with fluoride effects on *S. mutans* biofilms and dental caries. *J Dent Res.* 2005; 84(11): 1016–1020.
- [160] Koo H, Hayacibara M, Schobel B, Cury J, Rosalen P, Park Y, Vacca-Smith AM, Bowen WH. Inhibition of *Streptococcus mutans* biofilm accumulation and polysaccharide production by apigenin and tt-farnesol. J Antimicrob Chemother. 2003; 52(5): 782–

789.

- [161] Ohemeng K, Schwender C, Fu K, Barrett J. DNA gyrase inhibitory and antibacterial activity of some flavones (1). *Bioorganic Med Chem Lett.* 1993; 3(2): 225–230.
- [162] Wang M, Firrman J, Zhang L, Arango-Argoty G, Tomasula P, Liu L, Xiao W, Yam K. Apigenin impacts the growth of the gut microbiota and alters the gene expression of *Enterococcus. Molecules.* 2017; 22(8): 1–22.
- [163] Lv X, Qiu M, Chen D, Zheng N, Jin Y, Wu Z. Apigenin inhibits enterovirus 71 replication through suppressing viral IRES activity and modulating cellular JNK pathway. *Antiviral Res.* 2014; 109: 30–41.
- [164] Shibata C, Ohno M, Otsuka M, Kishikawa T, Goto K, Muroyama R, Kato N, Yoshikawa T, Takata A, Koike K. The flavonoid apigenin inhibits hepatitis C virus replication by decreasing mature microRNA122 levels. *Virology*. 2014; 462-463: 42–48.
- [165] Qian S, Fan W, Qian P, Zhang D, Wei Y, Chen H, Li X. Apigenin restricts FMDV infection and inhibits viral IRES driven translational activity. *Viruses*. 2015; 7(4): 1613–1626.
- [166] Hakobyan A, Arabyan E, Avetisyan A, Abroyan L, Hakobyan L, Zakaryan H. Apigenin inhibits African swine fever virus infection *in vitro*. *Arch Virol*. 2016; 161(12): 3445–3453.
- [167] Lee H, Woo ER, Lee DG. Apigenin induces cell shrinkage in *Candida albicans* by membrane perturbation. *FEMS Yeast Res.* 2018; 18(1): 1–9.
- [168] Fonseca-Silva F, Canto-Cavalheiro MM, Menna-Barreto RF, Almeida-Amaral EE. Effect of apigenin on *Leishmania amazonensis* is associated with reactive oxygen species production followed by mitochondrial dysfunction. J Nat Prod. 2015; 78(4): 880–884.
- [169] Ren B, Qin W, Wu F, Wang S, Pan C, Wang L, Zeng B, Ma S, Liang J. Apigenin and naringenin regulate glucose and lipid metabolism, and ameliorate vascular dysfunction in type 2 diabetic rats. *Eur J Pharmacol.* 2016; 773: 13–23.
- [170] Mao XY, Yu J, Liu ZQ, Zhou HH. Apigenin attenuates diabetes-associated cognitive decline in rats via suppressing oxidative stress and nitric oxide synthase pathway. *Int J Clin Exp Med.* 2015; 8(9):

15506-15513.

- [171] Eisenbarth GS. Update in type 1 diabetes. J Clin Endocrinol Metab. 2007; 92(7): 2403– 2407.
- [172] Johnson AE, Gordon C, Palmer RG, Bacon PA. The prevalence and incidence of systemic lupus erythematosus in Birmingham, England. Arthritis Rheum. 1995; 38(4): 551– 558.
- [173] Kang HK, Ecklund D, Liu M, Datta SK. Apigenin, a non-mutagenic dietary flavonoid, suppresses lupus by inhibiting autoantigen presentation for expansion of autoreactive Th1 and Th17 cells. *Arthritis Res Ther.* 2009; 11(2): 1–13.
- [174] Marshak-Rothstein A, Rifkin IR. Immunologically active autoantigens: the role of toll-like receptors in the development of chronic inflammatory disease. *Annu Rev Immunol.* 2007; 25: 419–441.
- [175] Verbeek R, van Tol EA, van Noort JM. Oral flavonoids delay recovery from experimental autoimmune encephalomyelitis in SJL mice. *Biochem Pharmacol.* 2005; 70(2): 220–228.
- [176] O'Connell RM, Taganov KD, Boldin MP, Cheng G, Baltimore D. MicroRNA-155 is induced during the macrophage inflammatory response. *Proc Natl Acad Sci.* 2007; 104(5): 1604–1609.
- [177] Maron BJ, Towbin JA, Thiene G, Antzelevitch C, Corrado D, Arnett D, Moss AJ, Seidman CE, Young JB. Contemporary definitions and classification of the cardiomyopathies: an American Heart Association scientific statement from the council on clinical cardiology, heart failure and transplantation committee; quality of care and outcomes research and functional genomics and translational biology interdisciplinary working groups; and council on epidemiology and prevention. Circulation. 2006; 113(14): 1807-1816.
- [178] Liu HJ, Fan YL, Liao HH, Liu Y, Chen S, Ma ZG, Zhang N, Yang Z, Deng W, Tang QZ. Apigenin alleviates STZ-induced diabetic cardiomyopathy. *Mol Cell Biochem*. 2017; 428(1-2): 9–21.
- [179] Mascaraque C, González R, Suárez MD, Zarzuelo A, de Medina FS, Martínez-Augustin O. Intestinal anti-inflammatory activity of apigenin K in two rat colitis models induced by trinitrobenzenesulfonic

acid and dextran sulphate sodium. *Br J Nutr*. 2015; 113(4): 618–626.

- [180] Rahimi R, Shams-Ardekani MR, Abdollahi M. A review of the efficacy of traditional Iranian medicine for inflammatory bowel disease. *World J Gastroenterol*. 2010; 16(36): 4504–4514.
- [181] Ghiu A, Schwiebs A, Radeke HH, Avram S, Zupko I, Bor A, Pavel IZ, Dehelean CA, Oprean C, Bojin F, Farcas C, Soica C, Duicu O, Danciu C. A comprehensive assessment of apigenin as an antiproliferative, proapoptotic, antiangiogenic and immunomodulatory phytocompound. *Nutrients*. 2019; 11(4): 1– 19.
- [182] Lefort ÉC, Blay J. Apigenin and its impact on gastrointestinal cancers. *Mol Nutr Food Res.* 2013; 57(1): 126–144.
- [183] Birt DF, Walker B, Tibbels MG, Bresnick E. Anti-mutagenesis and anti-promotion by apigenin, robinetin and indole-3-carbinol. *Carcinogenesis*. 1986; 7(6): 959–963.
- [184] Imran M, Aslam Gondal T, Atif M, Shahbaz M, Batool Qaisarani T, Hanif Mughal M, Salehi B, Martorell M, Sharifi-Rad J. Apigenin as an anticancer agent. *Phytother Res.* 2020; 34(8): 1812–1828.
- [185] Shukla S, MacLennan GT, Flask CA, Fu P, Mishra A, Resnick MI, Gupta S. Blockade of β -catenin signaling by plant flavonoid apigenin suppresses prostate carcinogenesis in TRAMP mice. *Cancer Res.* 2007; 67(14): 6925–6935.
- [186] Lee WJ, Chen WK, Wang CJ, Lin WL, Tseng TH. Apigenin inhibits HGF-promoted invasive growth and metastasis involving blocking PI3K/Akt pathway and β4 integrin function in MDA-MB-231 breast cancer cells. *Toxicol Appl Pharmacol.* 2008; 226(2): 178– 191.
- [187] Caltagirone S, Rossi C, Poggi A, Ranelletti FO, Natali PG, Brunetti M, Aiello FB, Piantelli M. Flavonoids apigenin and quercetin inhibit melanoma growth and metastatic potential. *Int J Cancer Res.* 2000; 87(4): 595–600.
- [188] Tseng TH, Chien MH, Lin WL, Wen YC, Chow JM, Chen CK, Kuo TC, Lee WJ. Inhibition of MDA-MB-231 breast cancer cell proliferation and tumor growth by apigenin through induction of G2/M arrest and histone H3 acetylation-mediated p21WAF1/CIP1 expression. *Environ Toxicol*.

2017; 32(2): 434-444.

- [189] Lee HH, Jung J, Moon A, Kang H, Cho H. Antitumor and anti-invasive effect of apigenin on human breast carcinoma through suppression of IL-6 expression. *Int J Mol Sci.* 2019; 20(13): 1–16.
- [190] Zhang S, Xu S, Duan H, Zhu Z, Yang Z, Cao J, Zhao Y, Huang Z, Wu Q, Duan J. A novel, highly-water-soluble apigenin derivative provides neuroprotection following ischemia in male rats by regulating the ERK/Nrf2/HO-1 pathway. *Eur J Pharmacol.* 2019; 855: 208–215.
- [191] Wang G, Li W, Lu X, Bao P, Zhao X. Luteolin ameliorates cardiac failure in type I diabetic cardiomyopathy. J Diabetes Complicat. 2012; 26(4): 259–265.
- [192] Kotanidou A, Xagorari A, Bagli E, Kitsanta P, Fotsis T, Papapetropoulos A, Roussos C. Luteolin reduces lipopolysaccharide-induced lethal toxicity and expression of proinflammatory molecules in mice. *Am J Respir Crit Care Med.* 2002; 165(6): 818–823.
- [193] Song YS, Park CM. Luteolin and luteolin-7-O-glucoside strengthen antioxidative potential through the modulation of Nrf2/MAPK mediated HO-1 signaling cascade in RAW 264.7 cells. *Food Chem Toxicol.* 2014; 65: 70–75.
- [194] Zhang T, Wu W, Li D, Xu T, Zhu H, Pan D, Zhu S, Liu Y. Anti-oxidant and antiapoptotic effects of luteolin on mice peritoneal macrophages stimulated by angiotensin II. *Int Immunopharmacol.* 2014; 20(2): 346–351.
- [195] Yang JT, Qian LB, Zhang FJ, Wang J, Ai H, Tang LH, Wang HP. Cardioprotective effects of luteolin on ischemia/reperfusion injury in diabetic rats are modulated by eNOS and the mitochondrial permeability transition pathway. *J Cardiovasc Pharmacol*. 2015; 65(4): 349–356.
- [196] Liao PH, Hung LM, Chen YH, Kuan YH, Zhang FBY, Lin RH, Shih HC, Tsai SK, Huang SS. Cardioprotective effects of luteolin during ischemia-reperfusion injury in rats. *Circ J.* 2011; 75(2): 443–450.
- [197] He D, Ma X, Chen Y, Cai Y, Ru X, Bruce IC, Xia Q, Shi G, Jin J. Luteolin inhibits pyrogallol-induced apoptosis through the extracellular signal-regulated kinase signaling pathway. *FEBS J.* 2012; 279(10):

1834–1843.

- [198] Liu L, Ma H, Yang N, Tang Y, Guo J, Tao W. A series of natural flavonoids as thrombin inhibitors: structure-activity relationships. *Thromb Res.* 2010; 126(5): e365–e378.
- [199] Choi JH, Kim YS, Shin CH, Lee HJ, Kim S. Antithrombotic activities of luteolin *in vitro* and *in vivo*. J Biochem Mol Toxicol. 2015; 29(12): 552–558.
- [200] Kim HJ, Lee W, Yun JM. Luteolin inhibits hyperglycemia-induced proinflammatory cytokine production and its epigenetic mechanism in human monocytes. *Phytother Res.* 2014; 28(9): 1383–1391.
- [201] Pace-Asciak CR, Hahn S, Diamandis EP, Soleas G, Goldberg DM. The red wine phenolics trans-resveratrol and quercetin block human platelet aggregation and eicosanoid synthesis: implications for protection against coronary heart disease. *Clin Chim Acta*. 1995; 235(2): 207–219.
- [202] Cyrus T, Witztum JL, Rader DJ, Tangirala R, Fazio S, Linton MF, Funk CD. Disruption of the 12/15-lipoxygenase gene diminishes atherosclerosis in apo E–deficient mice. J *Clin Invest.* 1999; 103(11): 1597–1604.
- [203] Juźwiak S, Wójcicki J, Mokrzycki K, Marchlewicz M, Białecka M, Wenda-Rózewicka L, Gawrońska-Szklarz B, Droździk M. Effect of quercetin on experimental hyperlipidemia and atherosclerosis in rabbits. *Pharmacol Rep.* 2005; 57(5): 604–609.
- [204] Kamada C, da Silva EL, Ohnishi-Kameyama M, Moon JH, Terao J. Attenuation of lipid peroxidation and hyperlipidemia by quercetin glucoside in the aorta of high cholesterol-fed rabbit. *Free Radic Res.* 2005; 39(2): 185–194.
- [205] Hayek T, Fuhrman B, Vaya J, Rosenblat M, Belinky P, Coleman R, Elis A, Aviram M. Reduced progression of atherosclerosis in apolipoprotein E–deficient mice following consumption of red wine, or its polyphenols quercetin or catechin, is associated with reduced susceptibility of LDL to oxidation and aggregation. *Arterioscler Thromb Vasc Biol.* 1997; 17(11): 2744–2752.
- [206] Kawai Y, Nishikawa T, Shiba Y, Saito S, Murota K, Shibata N, Kobayashi M, Kanayama M, Uchida K, Terao J. Macrophage as a target of quercetin glucuronides in human atherosclerotic

arteries implication in the anti-atherosclerotic mechanism of dietary flavonoids. *J Biol Chem.* 2008; 283(14): 9424–9434.

- [207] Enkhmaa B, Shiwaku K, Katsube T, Kitajima K, Anuurad E, Yamasaki M, Yamane Y. Mulberry (*Morus alba* L.) leaves and their major flavonol quercetin 3-(6malonylglucoside) attenuate atherosclerotic lesion development in LDL receptordeficient mice. J Nutr. 2005; 135(4): 729– 734.
- [208] Singh R, Singh B, Singh S, Kumar N, Kumar S, Arora S. Umbelliferone, an antioxidant isolated from *Acacia nilotica* (L.) Willd. Ex. Del. *Food Chem.* 2010; 120(3): 825–830.
- [209] Kim JS, Kim JC, Shim SH, Lee EJ, Jin WY, Bae K, Son KH, Kim HP, Kang SS, Chang HW. Chemical constituents of the root of *Dystaenia takeshimana* and their antiinflammatory activity. *Arch Pharm Res.* 2006; 29(8): 617–623.
- [210] Rezaee R, Behravan E, Behravan J, Soltani F, Naderi Y, Emami B, Iranshahi M. Antigenotoxic activities of the natural dietary coumarins umbelliferone, herniarin and 7-isopentenyloxy coumarin on human lymphocytes exposed to oxidative stress. *Drug Chem Toxicol*. 2014; 37(2): 144–148.
- [211] Lino C, Taveira M, Viana G, Matos F. Analgesic and antiinflammatory activities of *Justicia pectoralis* Jacq and its main constituents: coumarin and umbelliferone. *Phytother Res.* 1997; 11(3): 211–215.
- [212] Rauf A, Khan R, Khan H, Pervez S, Pirzada AS. *In vivo* antinociceptive and antiinflammatory activities of umbelliferone isolated from *Potentilla evestita*. *Nat Prod Res.* 2014; 28(17): 1371–1374.
- [213] Mazimba O. Umbelliferone: sources, chemistry and bioactivities review. *Bull Fac Pharm Cairo Univ.* 2017; 55(2): 223–232.
- [214] Ramu R, Shirahatti PS, Zameer F, Ranganatha LV, Prasad MN. Inhibitory effect of banana (*Musa* sp. var. Nanjangud rasa bale) flower extract and its constituents umbelliferone and lupeol on α -glucosidase, aldose reductase and glycation at multiple stages. *S Afr J Bot*. 2014; 95: 54–63.
- [215] Gao D, Zhang YL, Xu P, Lin YX, Yang FQ, Liu JH, Zhu HW, Xia ZN. In vitro evaluation of dual agonists for PPAR γ/β from the flower of *Edgeworthia gardneri* (wall.)

Meisn. J Ethnopharmacol. 2015; 162: 14–19.

- [216] Ramesh B, Pugalendi K. Impact of umbelliferone (7-hydroxycoumarin) on hepatic marker enzymes in streptozotocin diabetic rats. *Indian J Pharmacol.* 2006; 38(3): 209–210.
- [217] Sim MO, Ham JR, Lee HI, Seo KI, Lee MK. Long-term supplementation of umbelliferone and 4-methylumbelliferone alleviates high-fat diet induced hypertriglyceridemia and hyperglycemia in mice. *Chem Biol Interact.* 2014; 216: 9–16.
- [218] Yu SM, Hu DH, Zhang JJ. Umbelliferone exhibits anticancer activity via the induction of apoptosis and cell cycle arrest in HepG2 hepatocellular carcinoma cells. *Mol Med Rep.* 2015; 12(3): 3869–3873.
- [219] Kielbus M, Skalicka-Wozniak K, Grabarska A, Jeleniewicz W, Dmoszynska-Graniczka M, Marston A, Polberg K, Gawda P, Klatka J, Stepulak A. 7-Substituted coumarins inhibit proliferation and migration of laryngeal cancer cells *in vitro*. *Anticancer Res.* 2013; 33(10): 4347–4356.
- [220] Mousavi SH, Davari AS, Iranshahi M, Sabouri-Rad S, Tayarani Najaran Z. Comparative analysis of the cytotoxic effect of 7-prenyloxycoumarin compounds and herniarin on MCF-7 cell line. *Avicenna J Phytomed.* 2015; 5(6): 520–530.
- [221] Salehcheh M, Safari O, Khodayar MJ, Mojiri-Forushani H, Cheki M. The protective effect of herniarin on genotoxicity and apoptosis induced by cisplatin in bone marrow cells of rats. *Drug Chem Taxicol*. 2022; 45(4): 1470–1475.
- [222] Haghighitalab A, Matin MM, Bahrami AR, Iranshahi M, Haghighi F, Porsa H. Enhancement of cisplatin cytotoxicity in combination with herniarin in vitro. *Drug Chem Toxicol.* 2014; 37(2): 156–162.
- [223] Carvalho A, Silva D, Silva T, Scarcelli E, Manhani M. Evaluation of the antibacterial activity of ethanolic and cyclohexane extracts of chamomile flowers (*Matricaria chamomilla* L.). *Rev Bras Plant Med.* 2014; 16(3): 521–526.

Abbreviations

ABTS: 2,2-azino-bis-(3-ethylbenzthiazoline-6sulphonic acid); ABR: alveolar bone resorption; Ach: acetylcholine; ADP: adenosindiphosphat; ASFV: African swine fever virus; ALT: alanine aminotransferase; ALP: alkaline phosphatase; AST: aspartate aminotransferase; COX-2: cyclooxygenase-2; CTGF: myocardial connective tissue growth factor; DPPH: 2,2-Diphenyl-1picrylhydrazyl; eNOS: endothelial nitric oxide synthase; EV71: enterovirus 71; FMDV: footand-mouth disease virus; GAD: generalized anxiety disorder; GGT: gamma glutamyl transferase; GAE: gallic acid equivalent; GRAS: generally recognized as safe; GPx: glutathione peroxidase: GSH: glutathione; HAM-D: Hamilton depression rating; HCV: hepatitis C virus: HDL-CL: high-density lipoprotein cholesterol; 12-HETE: 12hydroxyeicosatetraenoate; HepG-2: human hepatoma cancer cells; His: histidine; HO-1: oxygenase-1; HPLC/NMR: heme highperformance liquid chromatography-nuclear HPLC-MS: magnetic resonance; highperformance liquid chromatography-mass spectrometry; hs-CRP: high sensitive-C reactive protein; ICAM: intracellular cell adhesion molecule; iNOS: inducible nitric oxide synthase; IFN: interferon; IL: interleukin; JNK: c-Jun Nterminal kinase; LDL: low density lipoprotein; 5-LOX: 5-lipoxygenase; MAPK: mitogen activated protein kinase; MIC: minimum inhibitory concentration; MMP-9: matrix metalloproteinase-9; MRSA: methicillin-resistant S. aureus; MDA: malondialdehyde; MS: multiple sclerosis; MPO: myeloperoxidase; NO: nitric oxide; NF-kB: nuclear factor kappa B; Nrf2: E2-related factor 2; premenstrual PMS: syndrome; **PPAR:** peroxisome proliferated activated receptor; QE: quercetin equivalent; RANKL/TRAP: receptoractivator of nuclear factor kappa beta/translocon associated protein; ROS: reactive oxygen specious; SLS: sodium lauryl sulfate; SOD: superoxide dismutase; STZ: streptozotocin; TG: triglyceride; TNF: tumor necrosing factor; TPN: total parental nutrition; VAS: visual analog scale; VCAM: vascular cell adhesion molecule; VEGF: Vascular endothelial growth factor