Research Journal of Pharmacognosy (RJP) 2(3), 2015: 9-16 Received: Mar 2015 Accepted: May 2015



Original article

Comparative evaluation of curcumin and curcumin loaded- dendrosome nanoparticle effects on the viability of SW480 colon carcinoma and Huh7 hepatoma cells

M. J. Dehghan Esmatabadi¹, M. K. Sarkandi², H. Motaleb Zadeh³, G. Khaledi⁴, M. Montazeri⁵, H. S. Zahed Shekarabi³, Y. Ayoubi Hormoz³, E. Ali Asgari⁶*

¹Department of Genetics, Faculty of Biological Sciences, Tarbiat Modares University, Tehran, Iran. ²Department of Microbiology, Faculty of Basic Sciences, Science and Research Branch, Islamic Azad University, Tehran, Iran

³Department of Genetics, Faculty of Basic Sciences, Science and Research Branch, Islamic Azad University, Tehran, Iran.

⁴Department of Biology, Faculty of Modern Sciences, Medical Branch of Tehran Islamic Azad University, Tehran, Iran.

⁵Department of Medical Biotechnology, Tabriz University of Medical Science, Tabriz, Iran. ⁶Department of Biology, Faculty of Basic Sciences, East Tehran Branch, Islamic Azad University, Tehran, Iran.

Abstract

Background and objectives: Colorectal cancer is the third most common cancer and a major cause of morbidity globally. Hepatocellular carcinoma is a leading cause of death in the world. About 80% of all anticancer drugs are somehow related to natural products. One of the most important of these natural compounds is curcumin, the main component of turmeric that has a wide range of pharmacological activities. Curcumin has been found to suppress cell proliferation and decrease cell viability in various types of cancer cells; however, owing to lack of aqueous solubility, curcumin has shown reduced bioavailability in studies. Recent studies have shown that new 400th generation of dendrosome nanoparticle can increase bioavailability of curcumin and thus enhance the cytotoxic properties. The aim of this study was to determine effectiveness of curcumin alone and in combination with 400th generation dendrosome nanoparticles (DNC) on cell viability rate in SW480 and Huh7 cells. **Methods:** SW480 and Huh7 cells were incubated with different concentrations of curcumin and DNC (0-50 μ M) for 24, 48 and 72 h. Then cytotoxicity was assessed by MTT assay and IC₅₀ was determined. **Results:** The results suggested that the concentration-dependent inhibitory effect of DNC was stronger than curcumin on SW480 and Huh7 cells. **Conclusion:** The results suggest DNC as a more effective herbal anticancer agent for colorectal and hepatocellular tumors.

Keywords: colorectal cancer, curcumin, dendrosomal curcumin, hepatocellular carcinoma, MTT assay

Available at: http://rjpharmacognosy.ir Copy right[©] 2014 by the Iranian Society of Pharmacognosy **Corresponding author: e.asgari@gmail.com, dnaresearchcenter@gmail.com,* Tel/Fax: +9891-25114523

Introduction

Colorectal cancer is the third most common cancer and a major cause of morbidity and mortality globally [1,2]. Although most Asian countries show very little incidence of colon cancer, it is the second leading cause of cancer deaths Western countries in [3]. Also. hepatocellular carcinoma (HCC) is one of the most prevalent cancers worldwide and its incidence is on the rise in developing countries [4]. Nowadays cancer takes place in result of different causes such as mutagenesis and carcinogen chemicals in the environment [5]. Most of mutagenic and carcinogen agents display their destructive effects through free radicals including reactive oxygen species (ROS). Antioxidants are able to reduce ROS; so, daily consumption of antioxidants enhances immunity of the body against free radicals production and serves as anticancer prevention [6]. Some fruits and vegetables are considered as the main anticancer foods, because of their abundant antioxidants such as phenols, vitamin C [7]. One of the most important of these natural compounds is curcumin (diferuloylmethane), the main component of turmeric that has a wide range of biological and pharmacological activities [8]. It has been used as a spice and a traditional medicine for many centuries in India and other Asian countries [9,10]. Chemo-preventive and therapeutic features of curcumin including anticancer, anti-inflammatory, antioxidant and antiproliferative activities were previously confirmed in in vitro and in vivo models [11]. The most important property of curcumin was observed to be its selective role in targeting cancer cells against normal cells [12,13]. In addition, curcumin was found to suppress cell proliferation and decrease cell viability rate in various types of cancer cells, including colon and liver [14-20]. However, owing to lack of aqueous solubility, curcumin has shown a reduced bioavailability in both in vitro and in vivo studies. The use of nanotechnology medicine in and more specifically drug delivery is set to spread rapidly. A multitude of substances are currently under investigation for preparation of nanoparticles for drug delivery, varying from biological substances like albumin, gelatine and phospholipids for liposomes, to substances with chemical nature like various polymers and solid metal containing nanoparticles. It is obvious that the potential interaction with tissues and cells, and toxicity, greatly depends on the actual composition of the nanoparticle formulation. To overcome the reduced bioavailability, various combinations of adjuvants, liposomes, dendrosomes and other substances have been used [21]. The anticancer properties of dendrosomal curcumin (DNC) as a coadministration of curcumin and dendrosome nanoparticle have been explored in mice models of fibrosarcoma and different cancers, and toxicological analysis have indicated severe side effects [22-25]. Particularly, findings of a recent study indicated curcumin loaded dedrosome nanoparticles to be efficient in terms of solubility, bioavailability, sustainability and thus enhanced anti-cancer properties [25]. Accordingly in this study, to assess the effects of these drugs alone and in combination with carrier, we compared the efficacy of curcumin-400th generation loaded dendrosome nanoparticles (DNC) versus free curcumin on cell viability rate in SW480 (colorectal cancer) and Huh7 (hepatocellular carcinoma) cells.

Experimental

Cell lines and chemicals

The human colon cancer SW480 and hepatocellular carcinoma Huh7 cell lines were obtained from Pasteur Institute of Iran (Tehran, Iran) and cultured in RPMI 1640 medium (Invitrogen) supplemented with 10 % fetal bovine serum (Invitrogen) and 1% penicillinstreptomycin (Gibco, Scotland) in 5 % CO₂ at 37 °C. Curcumin was purchased from Sigma-Aldrich Company, USA. Dendrosome nanoparticle specified as Den O400, a nonionic biodegradable denderic glycol ester (MW: 590 Da, HLB: 12.5 Mh/M, hydroxyl value: 95 mg KOH/g and acid value: mg KOH/g) was a gift from Institute of Biochemistry and Biophysics, University of Tehran, Iran [26,27].

Dendrosomal curcumin (DNC) preparation

For DNC preparation, we used the optimized protocol in our lab as previously described [28]; briefly, different weight/weight ratios of dendrosome/curcumin ranging from 50:1 to 10:1

were examined before settling a suitable ratio of 25:1. Curcumin was dissolved in various amounts of dendrosome and checked for absorbance spectra by UV spectrophotometery (TECAN, Switzerland). Then, the appropriate mixture of dendrosome and curcumin was evaluated for excitation/emission value in comparison with curcumin dissolved in PBS and 1% methanol as control samples. The loading of dendrosome nanocarriers with curcumin molecules was performed using MaLing Gou et al protocol [29]. Briefly, curcumin and dendrosome were codissolved in 5 mL of acetone; this solution was added to 5 mL of PBS while stirring constantly. Then. acetone was evaporated in rotary evaporator. The curcumin/dendrosome micelle solution was sterilized using a 0.22 µm syringe filter (Millex-LG, Millipore Co., USA). Finally, the prepared DNC was stored in 4 °C in a light protected condition until used. For in vitro experiments, DNC was diluted in complete culture medium as mentioned for each assay. Just like our previous studies, the curcumin loading efficiency was very high (87%) [28].

Microculture tetrazolium (MTT) assay

Microculture tetrazolium (MTT) assay was used to assess the inhibitory effect of DNC, free curcumin and free carrier (dendrosome) on viability of SW480 and Huh7 cells. For this purpose, seeded cells were plated in 96-well plates at density of 5000 cells/100 µl PRMI in each well and then, exposed to either control or a serial dilution of DNC, free curcumin and dendrosome ranging from 0 to 50 µM for 24, 48 and 72 h (in previous studies it was found that curcumin decreased the proliferation of SW480 and Huh7 cells with an inhibitory rate of nearly 100 % after 30-40 µM curcumin treatment, as a result the above mentioned concentrations were used). Thereafter, the control medium and the media were replaced by MTT solution (0.5 mg/ mL) and after 3 h, with DMSO. The absorbance was recorded at 570 nm with an ELISA reader (BioTek, USA). The percentage of viable cells was calculated as: (%) = (OD exp/ OD con) \times 100, where OD exp and OD con are the optical of exposed and densities control cells. respectively [30]. The 50 % inhibitory concentration (IC₅₀) values of DNC on SW480 and Huh7 cells at different time intervals were

determined by GraphPad Prism 5 statistical package. The parameter C or IC_{50} was calculated according to the following equation: $Y = [a-d/1+(X+c^b)] + d [31]$.

Statistical Analysis

The results were expressed as mean \pm SD. All experiments were performed in triplicate. Statistical significance of difference throughout this study were calculated using a two way ANOVA analysis. *p* <0.05 were considered significant.

Results and Discussion

The inhibitory evaluation, which was carried out through MTT assay using various concentrations (0-50 μ M) of DNC, free curcumin and dendrosome at different time intervals (24, 48 and 72 h), demonstrated that DNC and curcumin inhibited cell viability of SW480 and Huh7 cells in a concentration and time dependent manner (*p*<0.001).

As shown in figures 1 (a, b, c) and 2 (a, b, c), treatment of the cell lines with DNC (0–50 μ M) after 24, 48 and 72 h significantly reduced cell viability of SW480 and Huh7 cells more than free curcumin. However, after DNC and free curcumin treatment at 24 and 72 h the cell viability of Huh7 cells were very close to each other after 30 μ M. Also, no inhibitory effect was observed for dendrosome alone in both cell lines. These findings demonstrated that dendrosome increased water solubility and entry of curcumin to cells without toxic effects correlated to dendrosomes as a carrier.

In order to evaluate the performance of curcumin versus DNC, we compared the IC_{50} of curcumin loaded on dendrosome (DNC) to curcumin in SW480 and Huh7 cells (figures 3 and 4).

Cell sensitivity to curcumin was greatly increased when they were treated with DNC in comparison to curcumin as characterized by a decrease of IC_{50} for SW480 (from 38.72 to 16.09 µM) and Huh7 (from 40.22 to 22.3 µM) after 24 h. Also, 48 and 72 h treatment confirmed the increased cell chemo-sensitivity of SW480 and Huh7 cells to DNC ccompared to curcumin (figures 3 and 4). As controls, drug-unloaded dendrosome nanoparticles did not show any significant cell toxicity when used at concentrations close to the IC_{50} of DNC. Furthermore, coadministration of





Figure 1. Effects of DNC, free curcumin and dendrosome on SW480 cell viability. Results are expressed as a percentage of viability compared to control and are presented as mean±SD from three independent experiments.



Figure 2. Effects of DNC, free curcumin and dendrosome on Huh7 cell viability. Results are expressed as a percentage of viability compared to control and are presented as mean±SD from three independent experiments.



Figure. 3 IC₅₀ values of DNC and free curcumin on SW480 cells at different time intervals. Data are presented as mean \pm SD from three independent experiments. Asterisks show significant differences between the different treatment times compared to 24 h (* *P*<0.05, ** *p*<0.01, and # indicates significant differences between DNC and free

curcumin at each time point (p < 0.05, p < 0.01).



Figure 4. IC₅₀ values of DNC and free curcumin on Huh7 cells at different time intervals.

Asterisks show significant differences between the different treatment times compared to 24 h (* p < 0.05, ** p < 0.01).

drug-unloaded dendrosome and curcumin did not increase the anti-tumor property of curcumin alone, underlining that the cytotoxic activity was linked to the encapsulation of curcumin into dendrosome nanoparticles.

Despite the tremendous progress that has been achieved in the field of cancer biology, the rate of mortality has not changed remarkably due to high toxicity to normal tissues and low efficacy of anti-cancer drugs [13,25,32-34]. Thus, interest is increasing in natural products for prevention of carcinogenesis. Curcumin has a wide range of pharmacological effects, such as inhibition of cell proliferation, cell cycle progression and apoptosis in various cancer cell lines, including colon and liver cancer. It has been suggested that curcumin does not induce side effects of other anti-cancer drugs in *in vivo* and *in vitro* studies [12,13].

The inhibitory role of curcumin on progression and invasiveness of cancer cells has been observed through different in vitro and in vivo studies [33,35-46]. Gavin P. Collett et al. suggested that curcumin impedes colon cancer growth by inducing of apoptosis [47]. Notarbartolo et al. found that curcumin exerted cell growth inhibitory and apoptotic effects, at least partly, due to free radical generation which mainly depends on the activation of apoptosis in liver cancer [48]. But its optimum potential is limited by its lack of solubility in aqueous solvents and poor oral bioavailability. Some studies have previously suggested and designed many nanocarrier formulations for increasing curcumin efficiency. Maling Gou et al. suggested curcumin encapsulated into monomethoxy poly (ethyleneglycol)-poly(ε-caprolactone) (MPEG-PCL) micelles that induced a stronger anticancer effect than free curcumin in colon cancer [29]. Anand *et al.* employed a polymer-based nanoparticle approach to improve the bioavailability and curcumin-loaded PLGA nanoparticles formulation had enhanced cellular uptake, and increased bioactivity In vitro and superior bioavailability in vivo over curcumin [19].

In this research we used a new formulation of 400th generation dendrosome nanoparticles as curcumin carrier that had not shown toxicity against different normal cells in our previous studies. Our results, in agreement with previous reports, have clearly demonstrated that sensitivity of SW480 and Huh7 cells to curcumin was substantially increased with curcumin loaded in dendrosome (DNC) compared to free curcumin. In other word, DNC showed a higher cytotoxic activity than free curcumin. As a result, might dendrosome increase solubility, bioavailability and sustainability of curcumin and thus enhance its cytotoxic properties.

Declaration of interest

The authors declare that there is no conflict of interest. The authors alone are responsible for the content of the paper.

References

- [1] Goldberg RM, Sargent DJ, Morton RF, Fuchs CS, Ramanathan RK, Williamson SK, Findlay BP, Pitot HC, Alberts SR. A randomized controlled trial of fluorouracil plus leucovorin, irinotecan, and oxaliplatin combinations in patients with previously untreated metastatic colorectal cancer. J Clin Oncol. 2004; 22(1): 23-30.
- [2] Jemal A. Cancer statistics. *CA Cancer J Clin*. 2009; 59(4): 225-249.
- [3] Giovannucci E. Diet, body weight, and colorectal cancer: a summary of the epidemiologic evidence. *J Womens Health* (*Larchmt*). 2003; 12(2): 173-182.
- [4] Ikeda K, Saitoh S, Koida I, Arase Y, Tsubota A, Chayama K, Kumada H, Kawanishi M. A multivariate analysis of risk factors for hepatocellular carcinogenesis: A prospective observation of 795 patients with viral and alcoholic cirrhosis. *Hepatology*. 1993; 18(1): 47-53.
- [5] Namiki M. Antioxidants/antimutagens in food. Crc Cr Rev Food Sci. 1990; 29(4): 273-300.
- [6] Clarkson PM, Thompson HS. Antioxidants: what role do they play in physical activity and health. *Am J Clin Nutr*. 2000; 72(2): 637-646.
- [7] Ghasemian A, Mehrabian S, Majd A. Peel extracts of two Iranian cultivars of pomegranate (*Punica granatum*) have antioxidant and antimutagenic activities. *Pak J Biol Sci*. 2006; 9(7): 1402-1405.
- [8] Sporn MB. The War on Cancer: A Review. *Ann Ny Acad Sci.* 1997; 833: 137-146.
- [9] Kuo ML, Huang TS, Lin JK. Curcumin, an antioxidant and anti-tumor promoter, induces apoptosis in human leukemia cells. *Biochim Biophys Acta*. 1996; 1317(2): 95-100.
- [10] Ammon HP, Wahl MA. Pharmacology of *Curcuma longa. Planta Med.* 1991; 57(1): 1-7.
- [11] Khor TO, Keum YS, Lin W, Kim JH, Hu R, Shen G, Xu C, Gopalakrishnan A, Reddy B, Zheng X, Conney AH, Kong AN. Combined inhibitory effects of curcumin and phenethyl

isothiocyanate on the growth of human PC-3 prostate xenografts in immunodeficient mice. *Cancer Res.* 2006; 66(2): 613-621.

- [12] Ravindran J, Prasad S, Aggarwal BB. Curcumin and cancer cells: how many ways can curry kill tumor cells selectively. AAPS J. 2009; 11(3): 495-510.
- [13] Das T, Sa G, Saha B, Das K. Multifocal signal modulation therapy of cancer: ancient weapon, modern targets. *Mol Cell Biochem*. 2010; 336(1-2): 85-95.
- [14] Mehta K, Pantazis P, McQueen T, Aggarwal BB. Antiproliferative effect of curcumin (diferuloylmethane) against human breast tumor cell lines. *Anticancer Drugs*. 1997; 8(5): 470-481.
- [15] Radhakrishna Pillai GSA, Hassanein TI, Chauhan DP, Carrier E. Induction of apoptosis in human lung cancer cells by curcumin. *Cancer Lett.* 2004; 208(2): 163-170.
- [16] Rashmi R, Kumar S, Karunagaran D. Ectopic expression of Bcl-XL or Ku70 protects human colon cancer cells (SW480) against curcumin-induced apoptosis while their down-regulation potentiates it. *Carcinogenesis*. 2004; 25(10): 1867-1877.
- [17] Johnson JJ, Mukhtar H. Curcumin for chemoprevention of colon cancer. *Cancer Lett.* 2007; 255(2): 170–181.
- [18] Kim K, Kim KH, Kim HY, Cho HK, Sakamoto N, Cheong J. Curcumin inhibits hepatitis C virus replication via suppressing the Akt-SREBP-1 pathway. *Febs Letters*. 2010; 584(4): 707-712.
- [19] Anand P, Nair HB, Sung B, Kunnumakkara AB, Yadav VR, Tekmal RR, Aggarwal BB. Design of curcumin-loaded PLGA nanoparticles formulation with enhanced cellular uptake, and increased bioactivity in vitro and superior bioavailability *in vivo*. *Biochem Pharmacol*. 2010; 79(3): 330-338.
- [20] Kim HJ, Park SY, Park OJ, Kim YM. Curcumin suppresses migration and proliferation of Hep3B hepatocarcinoma cells through inhibition of the Wnt signaling

pathway. Mol Med Rep. 2013; 8(1): 282-286.

- [21] Babaei E, Sadeghizadeh M, Hassan ZM, Feizi MAH, Najafi F, Hashemi SM. Dendrosomal curcumin significantly suppresses cancer cell proliferation *in vitro* and *in vivo*. *Int Immunopharmacol*. 2012; 12(1): 226-234.
- [22] Sadeghizadeh M, Ranjbar B, Damaghi M, Khaki L, Sarbolouki MN, Najafi F. Dendrosomes as novel gene porters-III. J Chemi Technol Biotechnol. 2008; 83(6): 912-920.
- [23] Sarbolouki MN, Sadeghizadeh M, Yaghoobi MM, Karami A, Lohrasbi T. Dendrosomes: a novel family of vehicles for transfection and therapy. *J Chem Technol Biotechnol*. 2000; 75(10): 919-922.
- [24] Tahmasebi Mirgani M, Isacchi B, Sadeghizadeh M, Marra F, Bilia AR, Mowla SJ, Najafi F, Babaei E. Dendrosomalcurcumin nano formulation downregulates pluripotency genes via miR-145 activation in U87MG glioblastoma cells. *Int J Nanomed*. 2013; 9: 403-417.
- [25] Gou M, Men K, Shi H, Xiang M, Zhang J, Song J, Long J, Wan Y, Luo F, Zhao X, Qian Z. Curcumin-loaded biodegradable polymeric micelles for colon cancer therapy *in vitro* and *in vivo*. *Nanoscale*. 2011; 3(4): 1558-1567.
- [26] Mosmann T. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. J Immunol Methods. 1983; 65(1-2): 55-63.
- [27] Sebaugh JL. Guidelines for accurate EC_{50}/IC_{50} estimation. *Pharm Stat.* 2011; 10(2): 128-134.
- [28] Kimelman D, Xu W. Beta-catenin destruction complex: insights and questions from a structural perspective. *Oncogene*. 2006; 25(57): 7482-7491.
- [29] Chen HW, Lee JY, Huang JY, Wang CC, Chen WJ, Su SF, Huang CW, Ho CC, Chen JJ, Tsai MF, Yu SL, Yang PC. Curcumin inhibits lung cancer cell invasion and metastasis through the tumor suppressor HLJ1. *Cancer Res.* 2008; 68(18): 7428-7438.

- [30] Yu W, Xu YC, Tao Y, He P, Li Y, Wu T, Zhu YP, Li J, Wu JX, Dai J. DcR3 regulates the growth and metastatic potential of SW480 colon cancer cells. *Oncol Rep.* 2013; 30(6): 2741-2748.
- [31] Bharat BA, Shishir S, Yasunari T. Curcumin suppresses the paclitaxel-induced nuclear factor metastasis of human breast cancer in nude mice kb pathway in breast cancer cells and inhibits lung. *Clin cancer Res.* 2005; 11(20): 7490-7498.
- [32] Aggarwal S, Ichikawa H, Takada Y, Sandur SK, Shishodia S, Aggarwal BB. Curcumin (diferuloylmethane) down-regulates expression of cell proliferation and antiapoptotic and metastatic gene products through suppression of IkappaBalpha kinase and Akt activation. *Mol Pharmacol.* 2006; 69(1): 195-206.
- [33] Hong JH, Ahn KS, Bae E, Jeon SS, Choi HY. The effects of curcumin on the invasiveness of prostate cancer *in vitro* and *in vivo*. Prostate Cancer Prostatic Dis. 2006; 9(2): 147-152.
- [34] Su CC, Lin JG, Li TM, Chung JG, Yang JS, Ip SW, Lin WC, Chen GW. Curcumin. induced apoptosis of human colon cancer Colo 205 cells through the production of ROS, Ca²⁺and the activation of caspase-3. *Anticancer Res.* 2006; 26(6B): 4379-4389.
- [35] Wang X, Wang Q, Ives KL, Evers BM. Curcumin Inhibits neurotensin-mediated interleukin-8 production and migration of HCT116 human colon cancer cells. *Clin Cancer Res.* 2006; 12(18): 5346-5355.
- [36] Lin YG, Kunnumakkara AB, Nair A, Merritt WM, Han LY, Armaiz-Pena GN, Kamat AA, Spannuth WA, Gershenson DM, Lutgendorf SK, Aggarwal BB, Sood AK.Curcumin inhibits tumor growth and angiogenesis in ovarian carcinoma by targeting the nuclear factor-kappaB pathway. *Clin Cancer Res.* 2007; 13(11): 3423-3430.
- [37] Kim HI, Huang H, Cheepala S, Huang S, Chung J. Curcumin inhibition of integrin (alpha6beta4)-dependent breast cancer cell

motility and invasion. *Cancer Prev Res.* 2008; 1(5): 385-391.

- [38] Cai XZ, Wang J, Li XD, Wang GL, Liu FN, Cheng MS, Li F. Curcumin suppresses proliferation and invasion in human gastric cancer cells by downregulation of PAK1 activity and cyclin D1 expression. *Cancer Biol Ther.* 2009; 8(14): 1360-1368.
- [39] Herman JG, Stadelman HL, Roselli CE. Curcumin blocks CCL2 induced adhesion, motility and invasion, in part, through downregulation of CCL2 expression and proteolytic activity. *Int J Oncol.* 2009; 34(5): 1319-1327.
- [40] Bangaru ML, Chen S, Woodliff J, Kansra S. Curcumin (diferuloylmethane) induces apoptosis and blocks migration of human medulloblastoma cells. *Anticancer Res.* 2010; 30(2): 499-504.
- [41] Ibrahim A, El-Meligy A, Fetaih H, Dessouki A, Stoica G, Barhoumi R. Effect of curcumin and meriva on the lung metastasis

of murine mammary gland adenocarcinoma. *In Vivo*. 2010; 24(4): 401-408.

- [42] Mudduluru G, George-William JN, Muppala S, Asangani IA, Kumarswamy R, Nelson LD, Allgayer H. Curcumin regulates miR-21 expression and inhibits invasion and metastasis in colorectal cancer. *Bioscience Rep.* 2011; 31(3): 185-197.
- [43] Collett GP, Campbell FC. Curcumin induces c-jun N-terminal kinase-dependent apoptosis in HCT116 human colon cancer cells. *Carcinogenesis*. 2004; 25(11): 2183-2189.
- [44] Notarbartolo M, Poma P, Perri D, Dusonchet L, Cervello M, D'Alessandro N. Antitumor effects of curcumin, alone or in combination with cisplatin or doxorubicin, on human hepatic cancer cells. Analysis of their possible relationship to changes in NF-kB activation levels and in IAP gene expression. *Cancer Lett.* 2005; 224(1): 53-65.